

1 **Soil contaminations in landfill: a case study of the landfill** 2 **in Czech Republic**

3
4 **D. Adamcová¹, M. D. Vaverková¹, S. Bartoň², Z. Havlíček³ and E. Břoušková³**

5 [1]{Department of Applied and Landscape Ecology, Faculty of Agronomy, Mendel
6 University in Brno, Zemědělská 1, 613 00 Brno, Czech Republic}

7 [2]{Department of Technology and Automobile Transport, Faculty of Agronomy, Mendel
8 University in Brno, Zemědělská 1, 613 00 Brno, Czech Republic}

9 [3]{Department of Morphology, Physiology and Animal Genetics, Faculty of Agronomy,
10 Mendel University in Brno, Zemědělská 1, 613 00 Brno, Czech Republic}

11 Correspondence to: M.D. Vaverková (magda.vaverkova@uake.cz)

12 13 **Abstract**

14 Phytotoxicity test was determined to assess ecotoxicity of landfill soil. *Sinapis alba* L. was
15 used as heavy metals bioindicator. Soil samples 1 - 8, which were taken from the landfill
16 body, edge of the landfill body and its vicinity meet the limits for heavy metals Co, Cd, Pb,
17 and Zn specified in the applicable legislation. Hg and Mn threshold values are not established
18 in legislation, but values have been determined for the needs of the landfill operator. For
19 heavy metals Cr, Cu, and Ni sample 2 exceeded the threshold values, which attained the
20 highest values of all the samples tested for Cr, Cu and Ni. For Cr and Ni the values were
21 several times higher than values of the other samples. The second highest values for Cr, Cu,
22 and Ni showed sample 6 and 7. Both samples exceeded the set limits. An increase in plant
23 biomass was observed in plants growing on plates with soil samples, but no changes in
24 appearance, slow growth or necrotic lesions appeared. Ecotoxicity tests show that tested soils
25 (concentration of 50%) collected from the landfill body, edge of the landfill body and its
26 vicinity reach high percentage values of germination capacity of seeds of *Sinapis alba* L.
27 (101-137%). At a concentration of 25%, tested soil samples exhibit lower values of
28 germination capacity; in particular samples 3 to 8, yet the seed germination capacity in all 8
29 samples of tested soils range between 86 and 137%.

1 **1 Introduction**

2 Land degradation caused by human activities creates significant adverse effects on the
3 environments and ecosystems worldwide (Thomaz and Luiz, 2012; Bai et al., 2013; Li et al.,
4 2013; Chen et al., 2015) and solid waste is an important and emerging environmental
5 problem. It was estimated that 0.5–4.5 kg per person per day of solid waste is produced in
6 different regions of the world (Bakare et al., 2005; Swati et al., 2014). The most common
7 ways to manage such waste disposal are landfills and incinerators. Actually up to 95% total
8 municipal solid waste (MSW) collected is disposed of in landfills worldwide (El-Fadel et al.,
9 1997; Swati et al., 2014) and landfilling is the major MSW disposal method used in modern
10 cities (Wong et al., 2015). Landfills were thought to be the safe disposal method of MSW but
11 it is true only for properly engineered landfill sites. An engineered landfill site allows final
12 disposal of solid waste in a secure manner by minimizing the impacts on the environment as
13 modern landfills are often lined with layers of absorbent material and sheets of plastic to keep
14 pollutants from leaking into the soil and water (Swati et al., 2014).

15 The improper management of waste disposal raises public concern over potential harmful
16 effects to local communities and the environment. These concerns probably become more
17 pragmatic when recent intensive studies demonstrated increased human health risk caused by
18 exposure to toxic chemicals, such as dioxins and related compounds, and heavy metals in
19 these dumping sites (Agusa et al., 2003, Minh et al., 2003). Landfills containing hazardous
20 materials are under critical observation today for potential hazards, resulting in the need for
21 thorough risk analyses along with the soil and groundwater that have been contaminated with
22 chemicals leaching from landfills. Several reports have been published which are documented
23 on the leachate characterization and its effect on groundwater pollution (Boels and Fleming,
24 1993) but little information is available on the effect of landfills on the soil contamination
25 (Hernández et al., 1996) and its toxicological effects.

26 Soil pollution by heavy metals has become a critical environmental concern due to its
27 potential adverse ecological effects. Heavy metals occur naturally at low concentrations in
28 soils. However, they are considered as soil contaminants due to their widespread occurrence,
29 acute and chronic toxicity (Youn-Joo, 2014).

30 More recently high concentrations of heavy metal(loid)s, such as As, Cd, Cu, Pb, and Zn in
31 soils have often been reported in number of countries. For example, significant adverse
32 impacts of As on human health have been recorded in Bangladesh, India, and China and it is

1 claimed that millions of people are potentially at risk from As poisoning (Bhattacharya et al.,
2 2012). Similarly, Cd accumulation in the offal of grazing animals in New Zealand and
3 Australia made it unsuitable for human consumption and affected access of meat products to
4 overseas markets (Loganathan et al., 2008). Similarly, there have been concerns about urban
5 development of horticultural sites which contained toxic levels of metal(loid)s such as As, Cu,
6 and Pb in soils resulting from excessive use of fungicides and herbicides that are rich in these
7 metal(loid)s (Pietrzak and Uren, 2011).

8 Plants can be used as bioindicators for toxicity assessment in aquatic and terrestrial
9 ecosystems (Gorsuch et al., 1991). The present research was aimed at assessing the soil
10 pollution at the landfill site (in operation) and in the vicinity of a MSW landfill site. The main
11 objective of this study was characterization of soil samples issued from a landfill located near
12 Klatovy, in south-western Czech Republic, in relation to their content of heavy metals. The
13 other objective was to recommend some sensitive plant to assess phytotoxicity effect on one
14 vegetal specie. White mustard (*Sinapis alba* L.) was selected as the test plant species due to
15 their sensitivity to a wide range of contaminants. To assess phytotoxicity of landfill soil a
16 laboratory study was conducted.

17

18 **2 Material and methods**

19 **2.1 Site description**

20 The investigated landfill (Štěpánovice, 49°26'15.934"N, 13°16'55.352"E, ca. 405 m asl) is
21 located in Pilsen Region, western part of the Czech Republic. It started operating during 1996
22 with an authorized volume of 569000 m³, at the moment, it is being used to dispose mixed
23 municipal waste. The landfill is formed by three sub-landfills: landfill A (closed in 2003, area
24 8750 m²); landfill B (working from 2003, area 26000 m²); landfill C (that will work after
25 closing part B). The total volume of both (A, B) parts of the landfill is 289000 m³. Planned
26 service life of the facility is up to year 2018 (Vaverková and Adamcová, 2014a).

27 Every day, up to 37.5 tonnes of waste is authorized for landfilling after careful analysis: the
28 disposed waste includes municipal solid, non-hazardous wastes and the material for landfill
29 cover. Wastes may include scraps of paper, plastics and metals, packing, spent tires, textile
30 products, building materials, ashes from MSW incinerators, polluted terrain from
31 environment reclamation, etc. Particular details of waste composition, waste quantity stored

1 on landfill and landfill gas management are not presented in this article. Detailed information
2 and data were described in other articles (Vaverková and Adamcová, 2014a; Vaverková and
3 Adamcová, 2014b; Vaverková and Adamcová, 2014c).

4 The landfill site is located over an impermeable natural clay layer; bottom and side
5 boundaries may vary according to the period of cultivation, however they generally include
6 several protective layers, such as a compact clay layer (100 cm), geotextile membranes,
7 gravel (50 cm), geomembranes (2.5 mm) non-woven fabric (1200 g/m²), pulper products.

8 Landfill covers (top and side) are formed by a waste layer (terrain) to stabilize the surface,
9 drainage systems, compact clay (20 cm), soil bentonite and a vegetative soil layer (up to
10 100 cm). A grassy mantle and/or forestation with local vegetation will complete the recovery
11 of the environment after closing of each parcel. Systems for leachate treatment, and gas
12 recovery, collection and treatment are in operation. The landfill is situated in the north part of
13 widely opened valley directed towards W-E. The landfill is surrounded to the N and S by a
14 vegetation belt dominated by *Pinus sylvestris*. The hilly landscape in the western part of the
15 study area is used for agriculture, as well as the eastern lowland. The climate of the area is
16 typically inland, with mean annual rainfall over 582 mm and mean annual temperature of
17 8.0°C (Vaverková and Adamcová, 2015).

18

19 **2.2 Sample collection**

20 Soil samples were collected from landfill site at depth 10 cm (Fig. 1) in 2014. They were
21 collected in sterilized plastic containers. Freeze and grounded soil samples were homogenized
22 by sieving through a stainless steel 0.2-mm sieve, and stored in sealed containers at -4 °C
23 until analysis. The materials were analyzed for the content of heavy metals (Hg, Cd, Pb, Cu,
24 Zn, Co, Ni, Cr, Mn). The examined samples were brought to the testing laboratory
25 (Department of Chemistry and Biochemistry, Faculty of Agronomy, Mendel University in
26 Brno) for analyses.

27 Fig. 1 provides sampling points where soil samples were collected. In total 8 sampling points
28 were determined. Samples collected from sampling points 6, 7 and 8 were used as blind
29 samples. Samples from sampling points 4 and 5 were collected directly from the landfill body
30 and samples 1, 2 and 3 were taken from the edge of the landfill body. The allocations of
31 sampling sites were chosen on the basis of the authors` decision and on the grounds of mutual

1 comparison of the landfill body and its borders with the nearest vicinity of the landfill
2 (agriculturally utilized soil and forests).

3

4 **2.3 Sample processing and chemical analysis**

5 A microwave digestion (Ethos SEL, Milestone, Italy) was used for isolation of analytes from
6 solid samples. Soil samples were air-dried and sieved. A fraction < 2 mm was used for the
7 analysis. 200 ± 0.1 mg of dried and homogenized soil samples was used for partial digestion
8 in the microwave oven using 3 ml of concentrated HNO_3 and 9 ml of concentrated HCl at 200
9 °C and 1000W for 30 min. The soil digests were adjusted to the final volume of 25 ml with
10 deionized water.

11 Electrothermal atomic-absorption spectrometer (AAS ZEE nit 60, Analytic Jena, Germany
12 equipped with Zeeman correction) was used under the recommended conditions specified by
13 the manufacturer for determination Cd (228.8 nm), Pb (283.3 nm), Co (240.7 nm), Cr (359.3
14 nm). The wavelengths are given in parentheses. 1% Pd/Mg(NO_3)₂ was used as modifier.

15 Flame atomic-absorption spectrometer (AAS ZEE nit 60, Analytic Jena, Germany equipped
16 with Zeeman correction) was used under the recommended conditions specified by the
17 manufacturer for determination Cu (324.7 nm), Zn (213.8 nm), Ni (232.0 nm), Mn (279.5
18 nm). Acetylene-air flame was used for determination of analytes. The wavelengths are given
19 in parentheses.

20 Total mercury content in soil samples was measured by one purpose atomic absorption
21 spectrometer AMA 254 (Advanced Mercury Analyzer) controlled by WinAMA software
22 (both Altec, Prague, Czech Republic). The homogenized solid samples were weighted ($100 \pm$
23 0.1 mg) into pre-cleaned combustion boats and inserted into the AMA254 analyzer. During
24 analysis the sample was dried at 120 °C for 90 s and thermally decomposed at 550 °C for 180
25 s under an oxygen flow. Selectively trapped mercury was subsequently released from the gold
26 amalgamator by a brief heat-up and finally quantified (measuring cycle, 60 s) as Hg⁰ by the
27 cold-vapor AAS technique at 253.65 nm.

28 LODs (limit of detection) of methods were 0.1 µg/kg for Hg, 0.02 µg/kg for Cd, 0.38 µg/kg
29 for Pb, 3.08 µg/kg for Cu, 3.70 µg/kg for Zn, 4.92 µg/kg for Co, 9.00 µg/kg for Ni, 0.70

1 $\mu\text{g}/\text{kg}$ for Cr, and 12.10 $\mu\text{g}/\text{kg}$ for Mn. The results were in good agreement with the certified
2 values.

3 **2.4 Test plant species**

4 The test species were white mustard (*Sinapis alba* L.). They were selected because they are
5 known to be sensitive to board range of chemicals. White mustard is ideal for studying soils
6 and soil extracts (Gerencsér et al, 2010; OECD Guideline 208 for the Testing of Chemicals,
7 2003). Seeds were surface-sterilized by soaking for 2 min. in a commercial sodium
8 hypochlorite (2%) solution to which a few drops of Tween-20 had been added. Then they
9 were rinsed twice in sterile distilled water. Damage or empty seeds hulls were discarded.

10 **2.5 Phytotoxicity test**

11 The earthen pot experiment was performed under laboratory conditions ~~(Fig-2)~~. The earthen
12 pots (height of 10 cm and a diameter of 11 cm) were loosely filled with 200 g of medium,
13 than 100 seeds of white mustard were scattered on to the surface, covered with thin layer of
14 silica sand and covered with a glass plate (to avoid evaporation). The possible toxicological
15 effect was assessed according to CSN EN 13432 on growth of dicotyledonous plants. The
16 medium was specialized soil for germination and plant growth, enriched with soil samples
17 (25 %, 50 % w/w). Reference soil was composed from peat and silica sand. Plants were
18 grown under controlled conditions for 21 days. Humidity at level of 70 ± 25 % of water
19 absorption capacity was maintained to be constant. The toxicity tests were conducted at
20 ambient laboratory temperature of 22 ± 10 °C, continuous light was used. Values obtained
21 from two simultaneously conducted experiments were averaged and presented (germination
22 capacity, plant biomass).

23

24 **3 Results and discussion**

25 From the chemical analysis of solid samples with atomic absorption, the leaching values of
26 metals varied over a wide range as follows: Hg (0.0300 – 0.0663 mg/kg), Cd (0.0600 –
27 0.2044 mg/kg), Pb (2.5703 – 8.5287 mg/kg), Cu (32.43 – 51.24 mg/kg), Zn (25.67 – 41.97
28 mg/kg), Co (2.953 – 12.712 mg/kg), Ni (32.65 – 140.03 mg/kg), Cr (64.06 – 190.73 mg/kg) a
29 Mn (547.52 – 701.39 mg/kg). The **average** metal leaching values found in this study are
30 shown in Table 1.

1 Fig. 23 shows the graphically evaluated results of heavy metal content in individual soil
2 samples, with marked limit values for those heavy metals for which the limits are established
3 in the Decree of the Ministry of the Environment of the Czech Republic No. 13/1994 Coll., as
4 amended, laying down the details of agricultural land fund protection.

5 For heavy metals Cd, Pb, Zn, and Co, none of the eight examined soil samples did exceed the
6 limits specified in the Decree, as shown by the charts listed on Figure 3. For Cu, Ni, and Cr
7 some of the collected soil samples exceeded the limits established in the relevant legislation.
8 For Cu, the maximum limit for this heavy metal is 50 mg/kg. Samples that exceeded the limit
9 for Cu were as follows: sample 2 (58.62 mg/kg), sample 6 (51.24 mg/kg), and sample 8
10 (50.20 mg/kg). For Ni, the maximum limit is set at 25 mg/kg. This threshold value was
11 exceeded by all 8 soil samples; the highest value was measured in sample 2 (140.03 mg/kg).
12 The maximum allowed value for Cr (40 mg/kg) was exceeded by all 8 samples, with sample 2
13 showing the highest value (190.73 mg/kg). No limit values are established for Mn and Hg
14 presence in the soil. After the levels of heavy metals in collected soil samples were
15 determined, the phytotoxicity was tested.

16 Fourteen days from the establishment of the experiment, sprouts and the number of growing
17 plants occurring in the earthen pots were counted. The data were plotted into tables and
18 photographs were taken to document the course of the experiment. Germinating capacity and
19 growth of white mustard is shown in Fig. 34. Twenty-one days from the establishment of the
20 experiment, the counting of sprouts and growing plants was repeated, the results were
21 recorded and photographs were taken.

22 Values were calculated from the obtained data (Table 2) and results were evaluated. The
23 number of sprouts (number of growing plants) occurring on samples of examined soil and on
24 the soil from the blank experiment was compared for all mixing ratios. Germinating capacity
25 was calculated as a percentage of the corresponding values obtained from soils in the blank
26 experiment.

27 Table 23 lists average values calculated from the results obtained after conducting the
28 experiment (see Table 2) as well as percentages of germination capacity for each sample of
29 examined soil.

30 Fig. 45 shows the percentage expression of germination capacity of seeds of white mustard
31 (25% share of soil of samples 1-8) after 14 days from the start of the experiment and after 21
32 days (end of the experiment).

1 Maximum germination capacity of seeds of white mustard at a concentration of 25% was
2 achieved for sample 1, both in the period of 14 days (139%) and after 21 days (137%). The
3 second highest value exhibited sample 2 (131 % after 14 days and 136% after 21 days). The
4 third highest values were measured for samples 3 and 8, where after 14 days the germination
5 capacity reached 106% and 111%, respectively; and after 21 days the germination capacity
6 was 110% and 107%, respectively.

7 Even sample 4 exhibits high values of germination capacity after 14 days (102%) and after 21
8 days (103%). The 100% germination capacity limit was approached also by sample 5 after the
9 period of 21 days when the seed germination capacity attained 100%. The lowest values of
10 germination capacity of white mustard seeds showed samples 6 and 7. Germination capacity
11 of sample 6 was 85% after 14 days and 91% after 21 days, and that of sample 7 was 80% after
12 14 days and 86% after 21 days.

13 Fig. 56 shows the percentage expression of germination capacity of seeds of white mustard
14 (50% share of soil of samples 1-8) after 14 days from the start of the experiment and after 21
15 days (end of the experiment).

16 At a concentration of 50%, all samples (Samples 1-8) reported seed germination capacity
17 values over 100%, once after 14 days and again after the 21 day period. The highest values of
18 germination capacity occurred in sample 1 after 14 days (138%) and after 21 days (133%).
19 The second highest germination capacity was observed in sample 5, where it reached 123%
20 after 14 days and 122% after 21 days. The third place in germination capacity of white
21 mustard seeds was occupied by sample 7 (122% after 14 days and 119% after 21 days).

22 An increase in plant biomass was observed in plants growing on plates with soil samples from
23 the landfill body and its vicinity, but no changes in appearance, slow growth or necrotic
24 lesions appeared. Ecotoxicity tests show that tested soils (at a concentration of 50%) collected
25 from the landfill body, edge of the landfill body and its vicinity reach high percentage values
26 of germination capacity of seeds of white mustard (101-137%) (Fig. 67). At a concentration
27 of 25%, tested soil samples exhibit lower values of germination capacity; in particular
28 samples 3 to 8, yet the seed germination capacity in all 8 samples of tested soils range
29 between 86 and 137%.

30 The analysis of the variance is listed in Table 34. P(ANOVA) was calculated using the *Maple*
31 software. P-value determines the significance level, where it is possible to reject the
32 hypothesis that both models used are equivalent. P-value is compared with a pre-chosen

1 constant (most commonly 0.05) and when it is smaller, the equivalence of the models is
2 rejected. Three cases where the assumption is that the behaviour of the samples is different
3 from the behaviour of the blanks by 5% significance are marked in Table 34. Four
4 measurements were provided for each sample – two concentrations and two germination
5 rates. 4 values of p are available for each sample. Not one sample can be discarded in most
6 cases, see Fig 78. The values of p factor (ANOVA) for germination after 14 days are plotted
7 on the x-axis of Fig 78, the values of p factor (ANOVA) for germination after 21 days are
8 plotted on the y-axis. The green area is the requirement for equivalent germination – H_0 for
9 samples and blanks positively satisfied on the standard range of significance 0,05 – 5%. The
10 pink areas indicate the failure to satisfy this condition for one of the germination rates (14 or
11 21) days. Sample 5 is located in this area, but it is just below the line for the 21-day
12 germination rate, it satisfies the 14-day germination rate. Sample 1 is also located in this area
13 for the germination rate of 21 days but only for 25% concentration. No samples are located in
14 the red area where hypothesis H_0 can be positively rejected. Due to this it is possible to
15 consider the assumption H_0 are not significantly affected by the landfill.

16

17 **4. Conclusions**

18 Phytotoxicity test was determined to assess ecotoxicity of landfill soil. Fourteen days after the
19 establishment of the experiment, sprouts and the number of growing plants occurring in the
20 earthen pots were counted. The data were plotted into tables and photographs were taken to
21 document the course of the experiment. Twenty-one days from the establishment of the
22 experiment, the counting of sprouts and growing plants was repeated, the results were
23 recorded and photographs were taken. Results were evaluated from the acquired data. The
24 number of sprouts (number of growing plants) on the soil samples and on the soil from the
25 blank experiment was compared for all mixing ratios. Germinating capacity was calculated as
26 a percentage share of corresponding values obtained from the soil in the blank experiment.
27 Results in the tables (germinating capacity of seeds) are mean values obtained from the
28 conducted experiment.

29 Plant growth test can be good protocol to assess the phytotoxicity of soil contaminated by
30 heavy metals. White mustard is sensitive plant that can be used as heavy metals bioindicator.
31 Soil samples 1 to 8, which were taken from the landfill body, edge of the landfill body and its
32 vicinity meet the limits for heavy metals Co, Cd, Pb, and Zn specified in the applicable

1 legislation. Hg and Mn threshold values are not established in legislation, but values have
2 been determined for the needs of the landfill operator. For heavy metals Cr, Cu, and Ni some
3 samples exceeded the threshold values, namely sample 2, which attained the highest values of
4 all the samples tested for Cr, Cu and Ni. For Cr and Ni the values were several times higher
5 than values of the other samples.

6 After sample 2, the second highest values for Cr, Cu, and Ni showed sample 6 and also
7 sample 7, this one particularly for Cr and Ni. Both of these samples exceeded the set limits,
8 but their measured values were not as high as in the case of sample 2.

9 An increase in plant biomass was observed in plants growing on plates with soil samples from
10 the landfill body and its vicinity, but no changes in appearance, slow growth or necrotic
11 lesions appeared. Ecotoxicity tests show that tested soils (at a concentration of 50%) collected
12 from the landfill body, edge of the landfill body and its vicinity reach high percentage values
13 of germination capacity of seeds of white mustard (101-137%). At a concentration of 25%,
14 tested soil samples exhibit lower values of germination capacity; in particular samples 3 to 8,
15 yet the seed germination capacity in all 8 samples of tested soils range between 86 and 137%.

16

17 **Author contribution**

18 D. Adamcová, M.D. Vaverková, Z. Havlíček and E. Břoušková designed the experiments and
19 D. Adamcová and M.D. Vaverková carried them out. S. Bartoň performed the analysis of the
20 variance. M.D. Vaverková prepared the manuscript with contributions from all co-authors.

21

22 **Acknowledgements**

23 This study was supported by the IGA – Internal Grant Agency Faculty of Agronomy
24 MENDELU No. IP 13/2015 “Evaluation of the hygienic quality of recycled manure solids
25 used for dairy cattle bedding”. We also thank the city of Klatovy and the Technical Services
26 of the city of Klatovy. We would like to express our great appreciation to the management of
27 the landfill Štěpánovice. Namely, we are very grateful to Ing. Vladimír Král, Ph.D. and his
28 colleagues for their assistance and their willingness to provide their time so generously.

29

1 **References**

- 2 Agusa, T., Kunito, T., Nakashima, E., Minh, T.B., Tanabe, S., Subramanian, A., and Viet,
3 P.H.: Preliminary on trace element contamination in dumping sites of municipal wastes in
4 India and Vietnam, *J. Phys IV (Proceedings)*, 107, 21-24, 2003, doi: 10.1051/jp4:20030233.
- 5 An, Y-A.: Soil ecotoxicity assessment using cadmium sensitive plants, *Environ. Poll*, 127, 21-
6 26, 2004, doi: 10.1016/S0269-7491(03)00263-X.
- 7 Bai, X.Y., Wang, S.J., and Xiong, K.N.: Assessing spatial–temporal evolution processes of
8 karst rocky desertification land: indications for restoration strategies. *Land Degrad. Dev.*, 24,
9 47-56, 2013, doi:10.1002/ldr.1102.
- 10 Bakare, A.A., Mosuro, A.A., and Osibanjo O.: An in vivo evaluation of induction of
11 abnormal sperm morphology in mice by landfill leachates, *Mutat. Res-Gen Tox En*, 582, 28-
12 34, 2005, doi: 10.1016/j.mrgentox.2004.12.007
- 13 Bhattacharya, S., Gupta, K., Debnath, S., Ghosh, U.C., Chattopadhyay, D., and
14 Mukhopadhyay, A.: Arsenic bioaccumulation in rice and edible plants and subsequent
15 transmission through food chain in Bengal basin: a review of the perspectives for
16 environmental health, *Toxicol. Environ Chem*, 94, 429-441, 2012, doi:
17 10.1080/02772248.2012.657200.
- 18 Boels, D., and Fleming, G.: Chemical time bombs from landfills: Appraisal and modelling,
19 *Land Degrad. Dev.*, 4, 99–405, 1993, doi: 10.1002/ldr.3400040425.
- 20 Chen, X.W., Tsz-Fung, Wong, J., Mo, W.Y., Man, Y.B., Wang-Wai, Ng. Ch., and Wong
21 M.H.: Ecological Performance of the Restored South East New Territories (SENT) Landfill in
22 Hong Kong (2000–2012), *Land Degrad. Dev.*, 2015, DOI: 10.1002/ldr.2366.
- 23 El-Fadel, M., Findikakis, A.N., and Leckie, J.O.: Environmental impacts of solid waste
24 landfilling, *J. Environ Manage*, 50, 1–25, 1997, doi: 10.1006/jema.1995.0131.
- 25 Gerencsér, G., Murányi, E., Szendi, K., and Varga, C.: Ecotoxicological studies on Hungarian
26 peloids (medicinal muds), *Appl. Clay Sci*, 50, 47-50, 2010, doi: 10.1016/j.clay.2010.06.022.
- 27 Gorsuch, J.W., Lower, W.R., Lewis, M.A., and Wang, W.: *Plants for Toxicity Assessment*
28 2:12-28, ASTM STP 1115. ASTM, Philadelphia, 1991.

- 1 Hernández, A.J., Adarve, M.J., and Pastor, J.: Some impacts of urban waste landfills on
2 Mediterranean soils, *Land Degrad. Dev.*, 9, 21–33, 1998.
- 3 Li, X.L., Gao, J., Brierley, G., Qiao, Y.M., Zhang, J., and Yang, Y.W.: Rangeland
4 degradation on the Qinghai–Tibet Plateau: implications for rehabilitation, *Land Degrad. Dev.*,
5 24, 72–80, 2013, doi: 10.1002/ldr.1108.
- 6 Loganathan, P., Hedley, M.J., and Grace, N.D.: Pasture soils contaminated with fertilizer
7 derived cadmium and fluoride: livestock effects, *Rev. Environ Contam T.*, 192, 29–66, 2008,
8 doi: 10.1007/978-0-387-71724-1_2.
- 9 Minh, N.H., Minh, T.B., Watanabe, M., Kunisue, T., Shinsuke, I., Tanabe, S., Sakai, S.,
10 Subramanian, A., Sasikumar, K., Viet, P.H., Tuyen, B.C., Tana, T.S., and Prudente, M.S.:
11 Open dumping site in Asian developing countries: a potential source of polychlorinated
12 dibenzo-p-dioxins and polychlorinated dibenzofurans, *Environ. Sci T*, 37, 1493–1502, 2003,
13 doi: 10.1021/es026078s.
- 14 OECD Guideline 208 for the Testing of Chemicals, 2003. Seedling Emergence and Seedling
15 Growth Test.
- 16 Pietrzak, U., and Uren, N.: Remedial options for copper-contaminated vineyard soils, *Soil.*
17 *Res*, 49, 44–55, 2011.
- 18 Swati, Ghosh, P., Tanay D. M., and Thaku, I.S.: In vitro toxicity evaluation of organic extract
19 of landfill soil and its detoxification by indigenous pyrene-degrading *Bacillus* sp. ISTPY1,
20 *Int. Biodeter Biodeg*, 90, 145–151, 2014, doi: 10.1002/ldr.2237.
- 21 Thomaz, E.L., and Luiz, J.C.: Soil loss, soil degradation and rehabilitation in a degraded land
22 area in Guarapuava (Brazil), *Land Degrad. Dev.*, 23, 72–81, 2012, doi: 10.1002/ldr.1052.
- 23 Vaverková, M.D., and Adamcová, D.: Can vegetation indicate a municipal solid waste
24 landfill's impact on the environment?, *Pol. J Environ Stud*, 2, 501-503, 2014a.
- 25 Vaverková, M.D., and Adamcová, D.: Heavy metals uptake by selected plant species in the
26 landfill area of Štěpánovice, Czech Republic, *Pol. J Environ Stud*, 23, 2265-2269, 2014b, doi:
27 10.15244/pjoes/26106.
- 28 Vaverková, M.D., and Adamcová, D. Evaluation of landfill pollution with special emphasis
29 on heavy metals. *J. Ecol Eng* 2, 1-6. 2014c, doi: 10.12911/22998993.1094972.

1 Vaverková, M.D., and Adamcová, D.: Case study of landfill reclamation at Czech landfill
2 site, Environ. Eng Manage J, Paper accepted for publication, 2015.

3 Wong, M.H., Chan, Y.S.G., Zhang, Ch., and Wang-Wai, Ng. Ch.: Comparison of pioneer and
4 native woodland species growing on top of an engineered landfill, Hong Kong: restoration
5 program, Land. Degrad Dev, DOI: 10.1002/ldr.2380

6 Table 1. Content of heavy metals in examined soil samples.

Sample	Hg (mg/kg)	Cd (mg/kg)	Pb (mg/kg)	Cu (mg/kg)	Zn (mg/kg)	Co (mg/kg)	Ni (mg/kg)	Cr (mg/kg)	Mn (mg/kg)
1	0.0300	0.0670	6.5554	34.07	25.67	4.888	39.13	69.99	689.55
2	0.0311	0.1446	2.5703	58.62	34.20	12.712	140.03	190.73	608.67
3	0.0343	0.1446	5.1769	39.37	31.51	2.953	32.65	65.92	547.52
4	0.0663	0.1576	8.5287	34.25	39.29	5.825	36.94	64.06	610.10
5	0.0403	0.1343	5.1095	32.43	33.93	10.284	33.11	70.10	584.58
6	0.0386	0.2044	5.4088	51.24	41.97	6.874	44.05	86.69	625.12
7	0.0459	0.0600	5.0800	43.80	32.10	5.375	42.76	74.85	661.00
8	0.0312	0.1471	4.1255	50.20	31.68	5.469	37.59	69.94	701.39

7

1 ~~Table 2. Results for germination capacity of seeds of white mustard for examined samples.~~

Sample	Summary—germination test	
	14 days	21 days
-		
1A-25	79	82
1B-25	91	95
1A-50	99	99
1B-50	100	100
2A-25	88	95
2B-25	72	81
2A-50	83	88
2B-50	86	88
3A-25	61	68
3B-25	68	74
3A-50	81	85
3B-50	91	93
4A-25	66	70
4B-25	59	63
4A-50	63	65
4B-50	92	96
5A-25	53	54
5B-25	68	75
5A-50	91	94
5B-50	86	89
6A-25	48	55

6B-25	56	62
6A-50	82	87
6B-50	62	65
7A-25	56	61
7B-25	41	50
7A-50	84	87
7B-50	91	92
8A-25	57	59
8B-25	78	79
8A-50	84	86
8B-50	85	89
Blank-I	69	72
Blank-II	75	78
Blank-III	70	73
Blank-IV	52	56

1 *A ——— 1 ——— performance, ——— B ——— second ——— performance

- 1 Table 23. Average values and percentages of germination capacity of seeds of white mustard
 2 for examined samples.

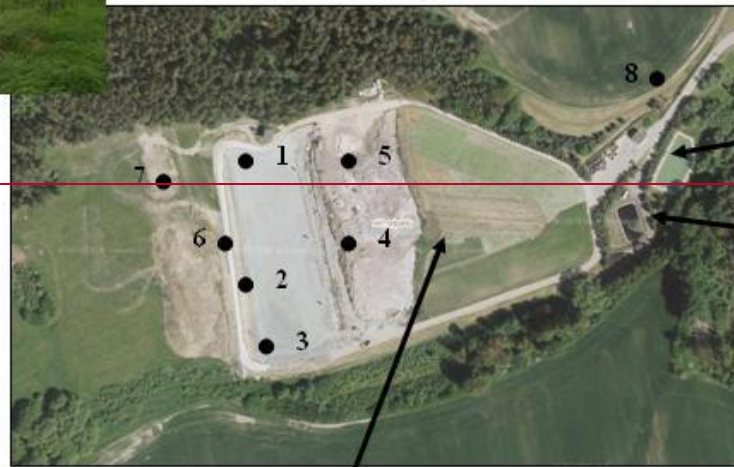
Sample - Mean	Summary - germination test		% number of seeds germinated	
	14 days	21 days	14 days	21 days
25%				
1	85	88.5	139	137
2	80	88	131	136
3	64.5	71	106	110
4	62.5	66.5	102	103
5	60.5	64.5	99	100
6	52	58.5	85	91
7	48.5	55.5	80	86
8	67.5	69	111	107
Blank	61	64.5	100	100
50%				
1	99.5	99.5	138	133
2	84.5	88	117	117
3	86	89	119	119
4	77.5	80.5	108	107
5	88.5	91.5	123	122
6	72	76	100	101
7	87.5	89.5	122	119
8	84.5	87.5	117	117
Blank	72	75	100	100

1 Table 34. Analysis of the variance.

		25%			
Sample A, B	14 days		21 days		
	Germ.	p(ANOVA)	Germ.	p(ANOVA)	
1	75, 91	0.091	82, 95	0.012	
2	88, 72	0.654	95, 81	0.076	
3	66, 68	0.811	68, 74	0.074	
4	66, 59	0.636	70, 63	0.398	
5	53, 68	0.533	54, 75	0.045	
6	48, 56	0.140	55, 62	0.601	
7	56, 41	0.110	61, 50	0.055	
8	57, 78	0.924	59, 79	0.075	
		50%			
Sample A, B	14 days		21 days		
	Germ.	p(ANOVA)	Germ	p(ANOVA)	
1	99, 100	0.084	99, 100	0.617	
2	83, 86	0.094	88, 88	0.063	
3	81, 91	0.874	85, 93	0.064	
4	63, 92	0.686	65, 96	0.417	
5	91, 86	0.535	94, 89	0.041	
6	82, 62	0.206	87, 65	0.559	
7	84, 91	0.146	87, 92	0.054	
8	84, 85	0.940	86, 89	0.070	
Blank	Germ. 14 days		Germ. 21 days		
I,II, III, IV	69, 75, 70, 52		72, 78, 73, 56		



Detail of the landfill



● Sampling points

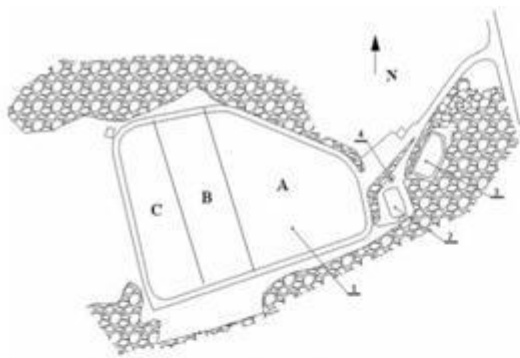
Landfill

Rainwater reservoir

Leachate pond

1

2



1 - landfill, 2 - detention receiver of leachate, 3 - rainwater reservoir, 4 - entrance gate

Section of the landfill



● Sampling points

Landfill

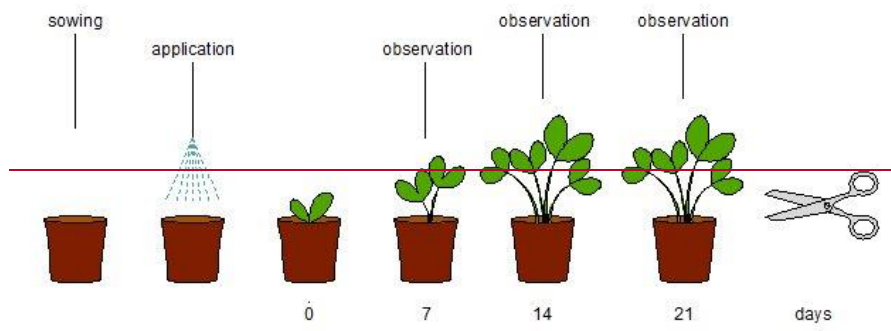
Rainwater reservoir

Leachate pond

3

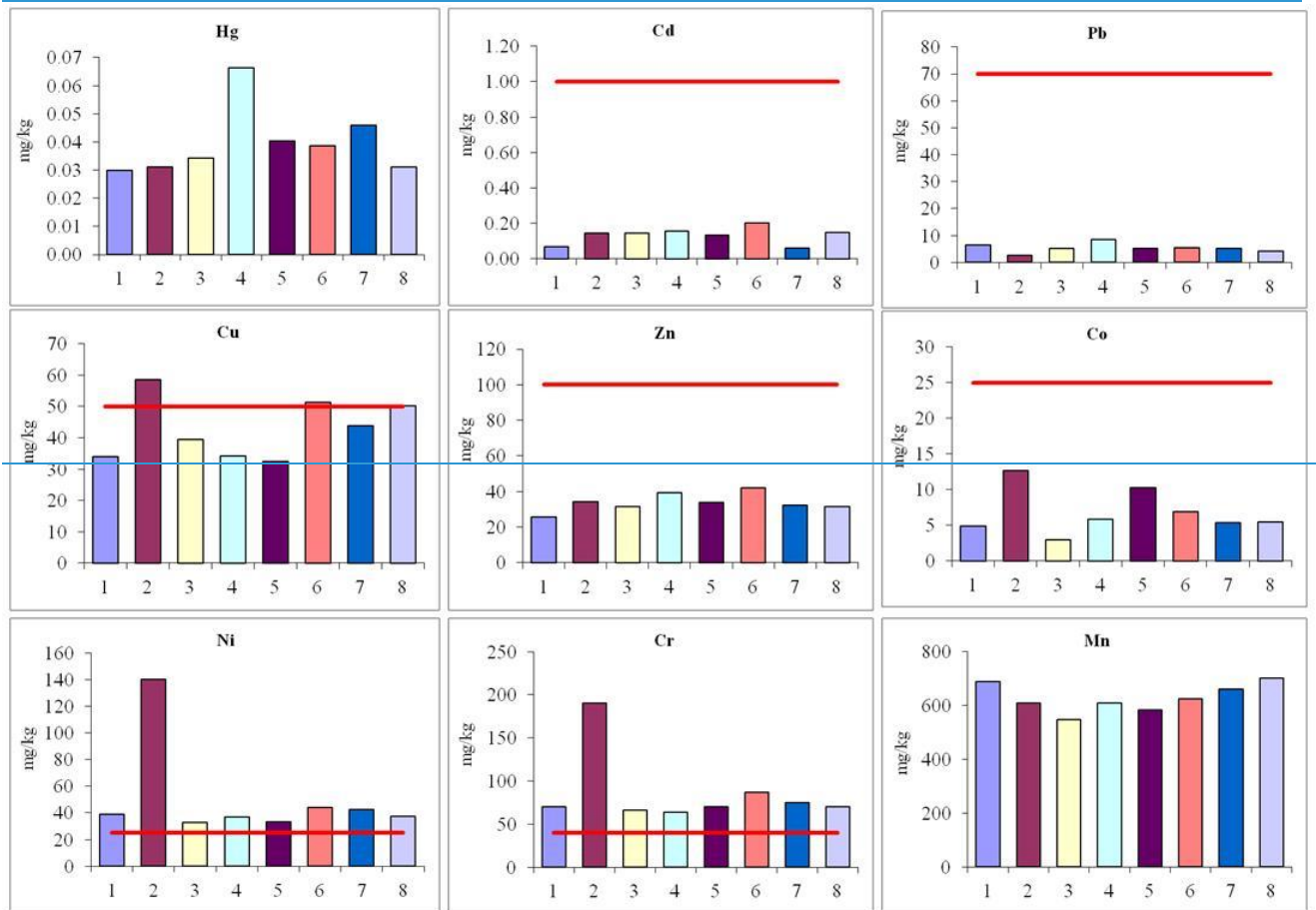
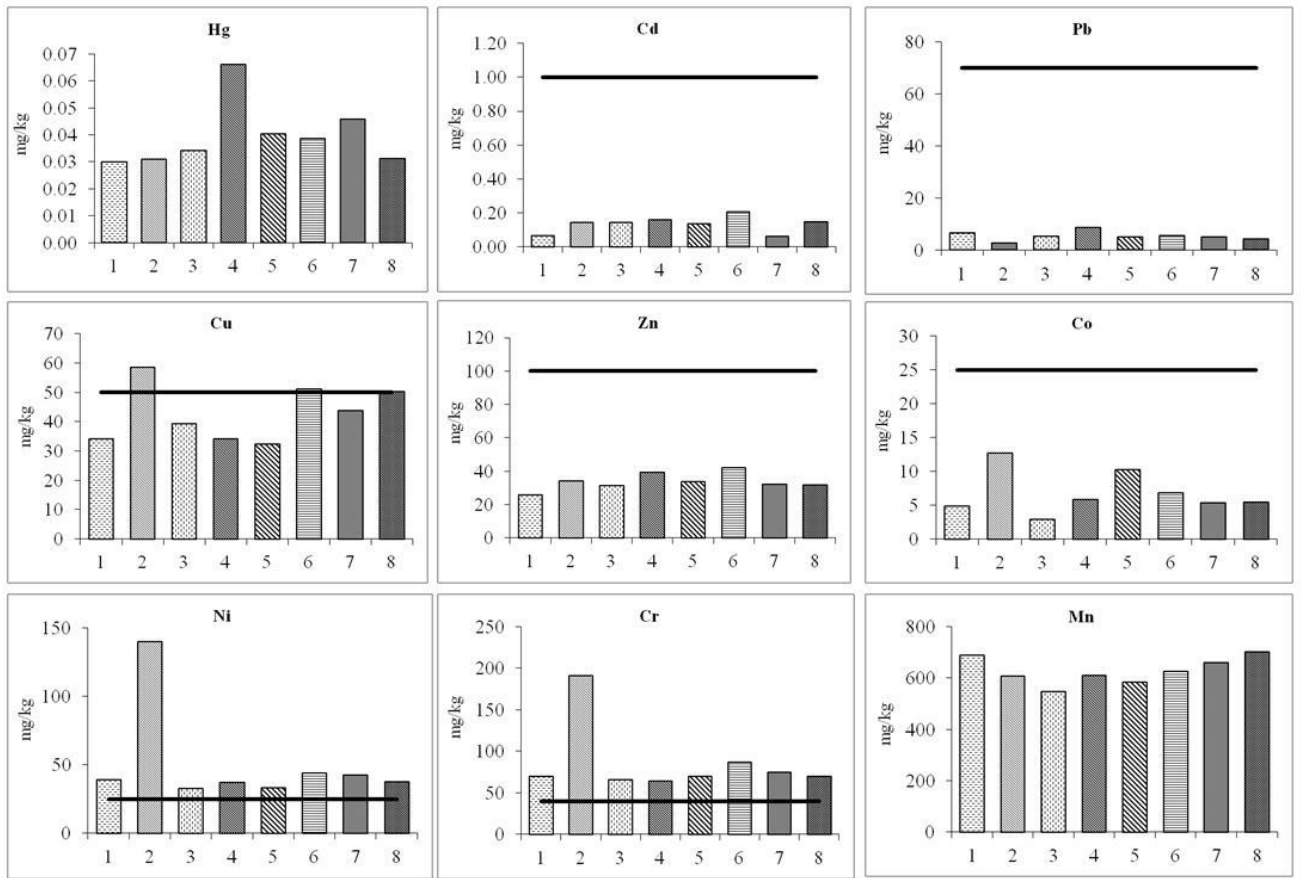
1 Figure 1. Map of Štěpánovice landfill and sampling points.

2



- 1
- 2
- 3
- 4

Figure 2. Layout of the phytotoxicity test.



1 Mn, Hg – no threshold values are set in the Decree No. 13/1994 Coll.

2

3 Figure 23. Content of heavy metals in examined soil samples with marked limit values set in

4 the Decree No. 13/1994 Coll.:-

5



1

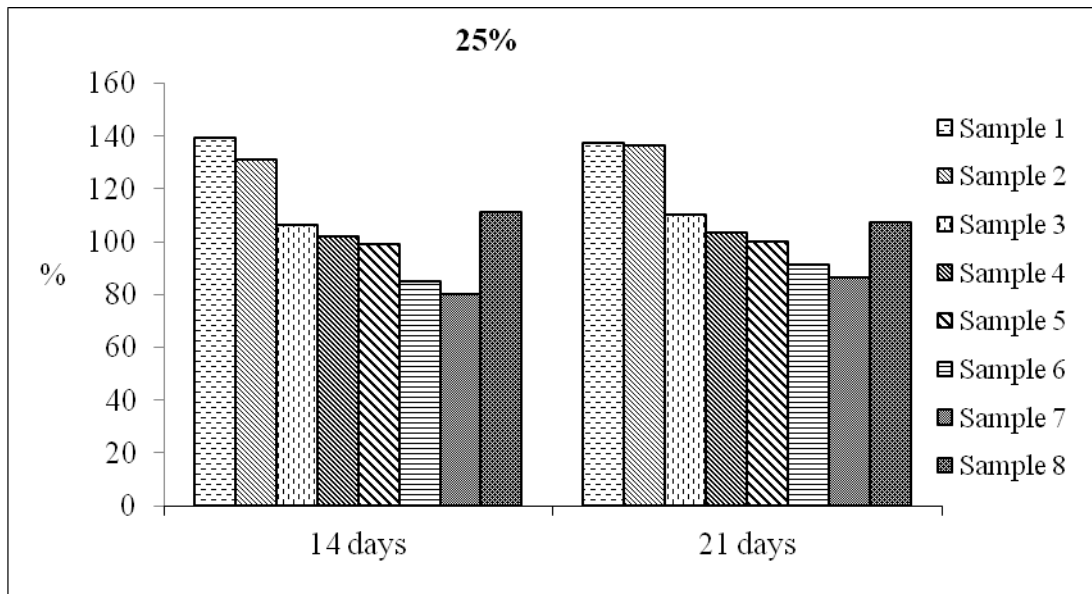


2

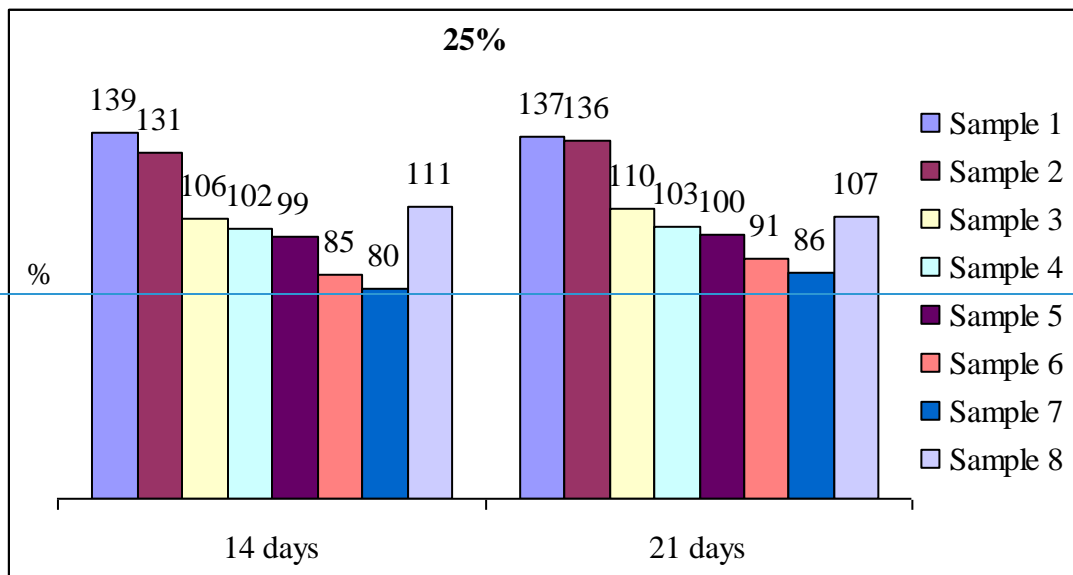
3

4 Figure 34. Samples of white mustard.

5



1



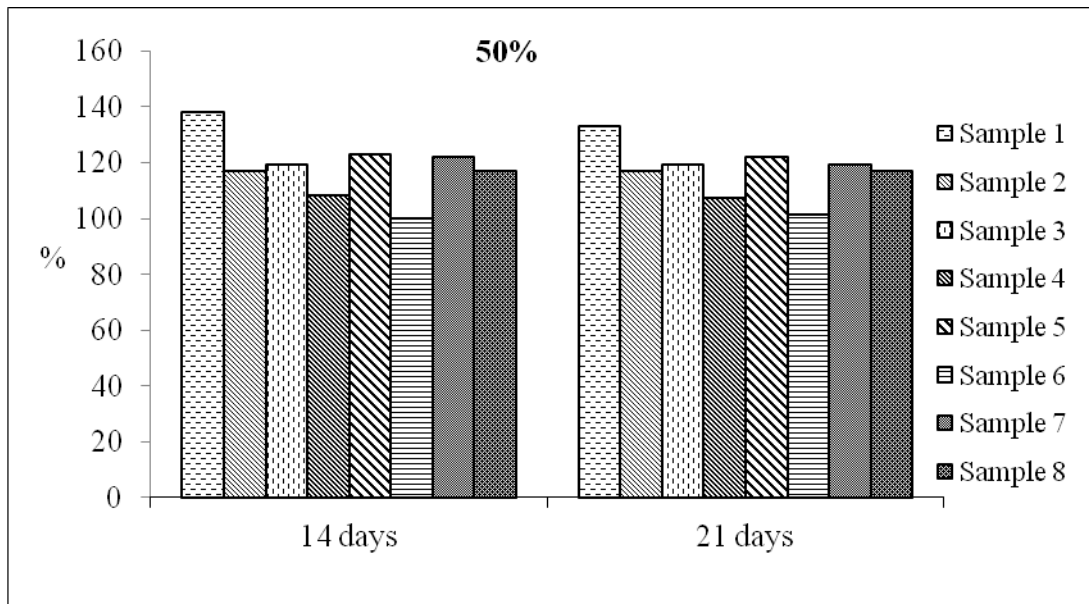
2

3

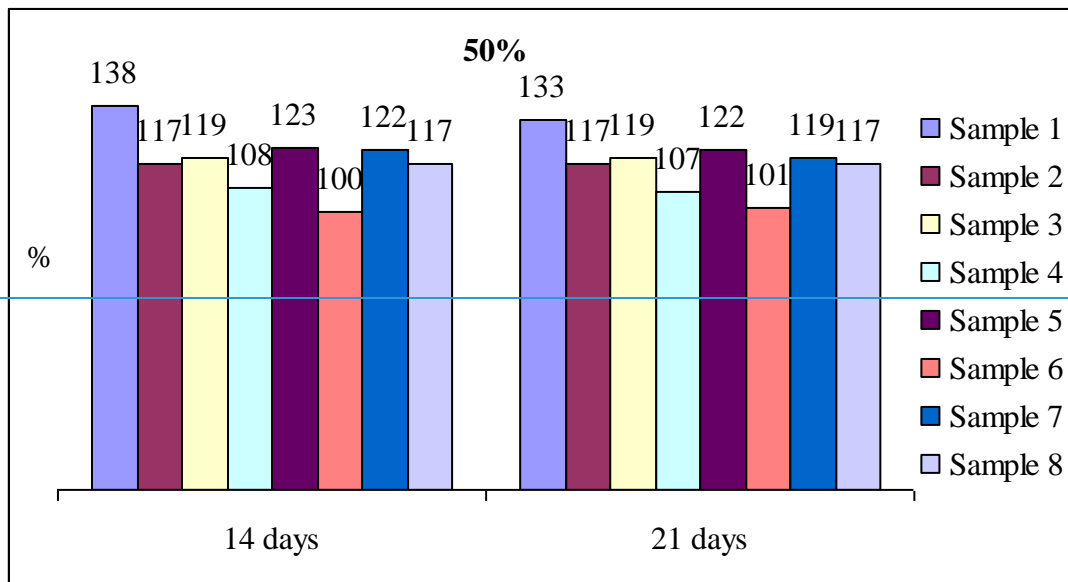
4

Figure 45. Comparison of the germination capacity at a concentration of 25%.

5



1

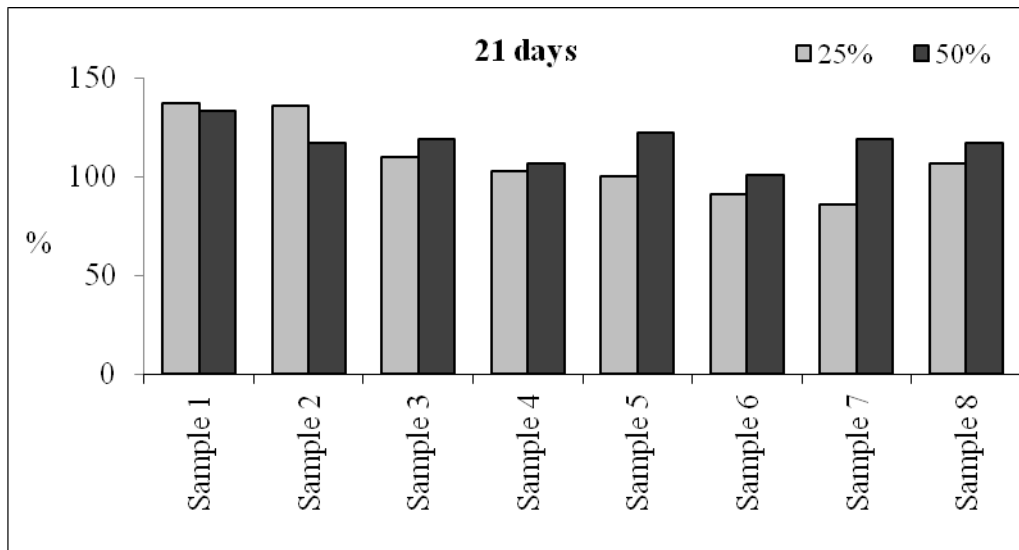


2

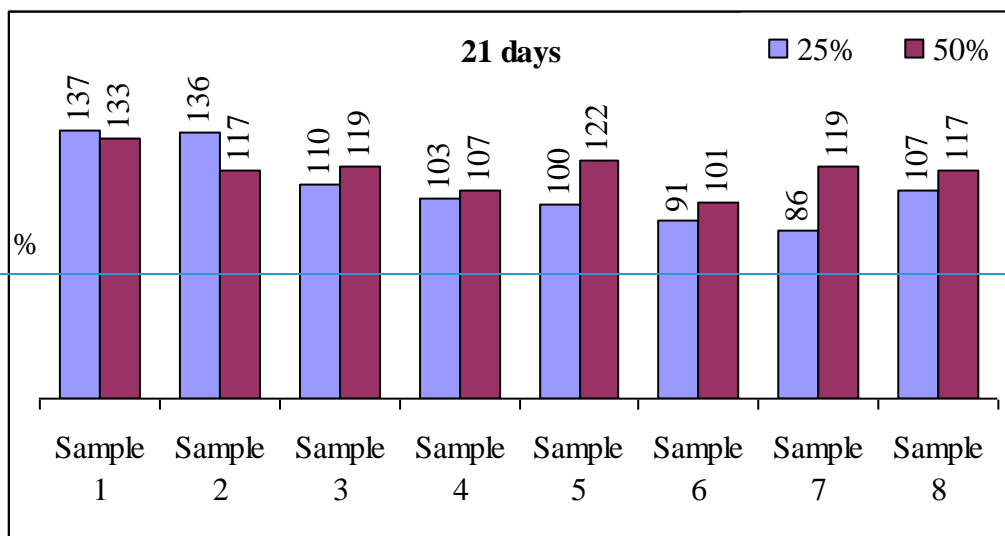
3

4 Figure 56. Comparison of the germination capacity of soil samples at a concentration of 50%.

5



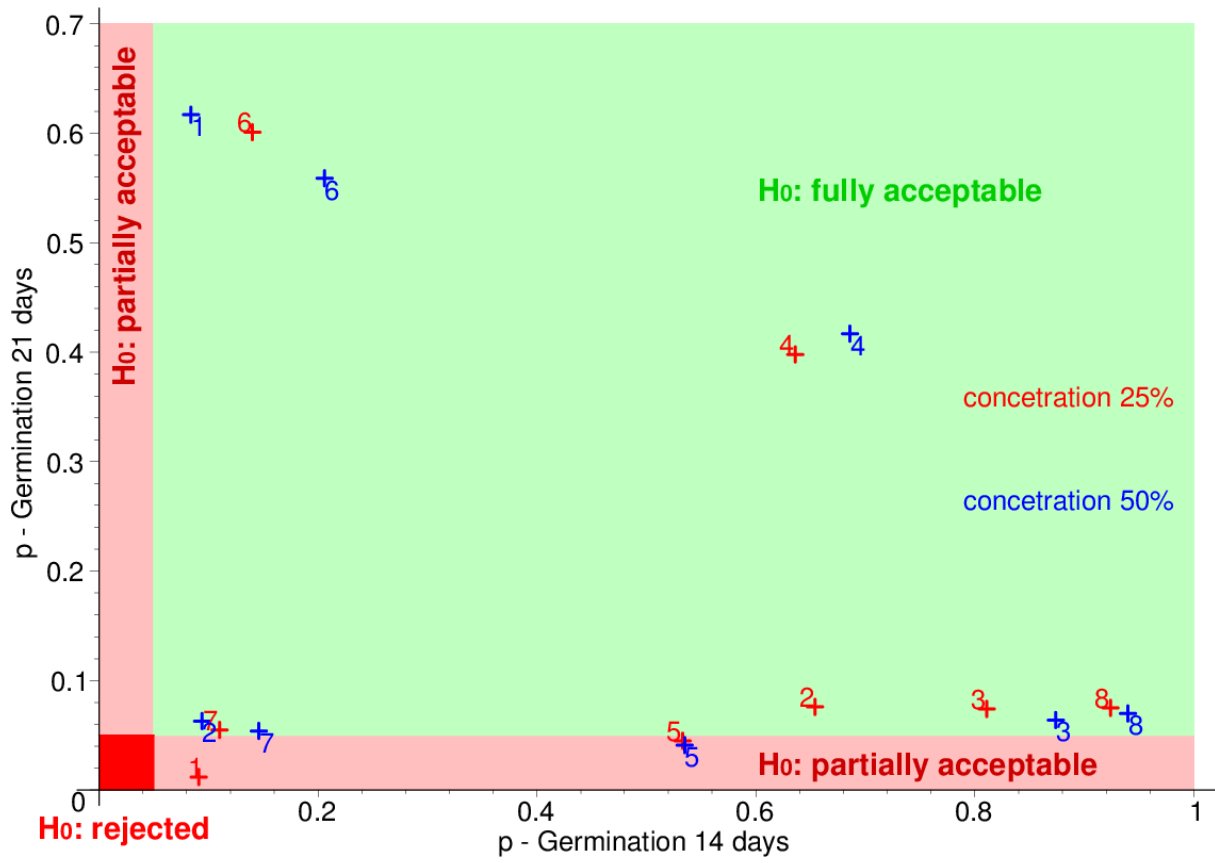
1



2

3

4 Figure 67. Results of germination capacity of white mustard seeds (at concentrations of 25%
5 and 50%).



1

2

3 Figure 78. The values of p factor (ANOVA)