



1 **Development of a composite soil degradation assessment**
2 **index for cocoa agroforests under tropical conditions of**
3 **southwest Nigeria**

4 Sunday Adenrele Adeniyi^{1,2}, Willem Petrus de Clercq³, and Adriaan van Niekerk^{1,4}

- 5
6 1. Department of Geography and Environmental Studies, Stellenbosch University, South Africa
7 2. Department of Geography, Osun State University, Nigeria
8 3. Department of Soil Science, Stellenbosch University, South Africa
9 4. School of Plant Biology, University of Western Australia, Australia

10 *Correspondence to:* Sunday Adenrele Adeniyi (releadegeography@yahoo.com)

11 **Abstract.** Cocoa agroforestry is a major landuse type in the tropical rainforest belt of West Africa, reportedly
12 associated with several ecological changes, including soil degradation. This study aims to develop a composite
13 soil degradation assessment index (CSDI) for determining the degradation level of cocoa soils under smallholder
14 agroforests of southwest Nigeria. Plots where natural forests have been converted to cocoa plantations of ages 1-
15 10 years, 11-40 years and 41-80 years, respectively representing young cocoa plantations (YCP), mature cocoa
16 plantations (MCP) and senescent cocoa plantations (SCP) were identified to represent the biological cycle of the
17 cocoa tree. Soil samples were collected at a depth of 0-20cm in each plot and analysed in terms of their physical,
18 chemical and biological properties. Factor analysis of soil data revealed four major interacting soil degradation
19 processes, decline in soil nutrient, loss of soil organic matter, increase in soil acidity and the breakdown of soil
20 textural characteristics over time. These processes were represented by eight soil properties (extractable zinc, silt,
21 SOM, CEC, available phosphorus, total porosity, pH, and clay). These soil properties were subjected to forward
22 stepwise discriminant analysis (STEPDA), and the result showed that four soil properties (extractable zinc; cation
23 exchange capacity; soil organic matter and clay) have the highest power to separate the studied soils into YCP,
24 MCP and SCP. In this way, we hope to have controlled sufficiently for redundancy in the final selection of soil
25 degradation indicators. Based on these four soil parameters, CSDI was developed and used to classify selected
26 cocoa soils into three (3) different classes of degradation. The results revealed that 65% of the selected cocoa
27 farms are moderately degraded, while 18% have a high degradation status. Finally, the value of the CSDI as an
28 objective index of soil degradation under cocoa agroforests was statistically validated.

29
30 **Keywords:** Smallholder cocoa agroforests, age-sequenced plantations, minimum data set, degradation indicators,
31 composite soil degradation assessment index, tropical conditions.

32
33
34



35 Introduction

36 Healthy soil is vital to successful agriculture and global food security (Virto, et al., 2014; Lal, 2015). Soil
37 performs several ecosystem functions such as carbon sequestration and regulation (Novara et al. 2011; Brevik et
38 al. 2015); buffering and filtering of pollutants (Keesstra et al. 2012); climate control through the regulation of C
39 and N fluxes (Brevik et al. 2015); and home for biodiversity (Schultecoo et al. 2015). Nonetheless, misuse of soils,
40 arising from intensive agricultural production and unsustainable land use practices have resulted in soil
41 degradation, particularly in developing countries with poor infrastructure and financial capacity to manage natural
42 resources (Teshahunegn, 2016). Statistics show that 500 million hectare (Mha) of land in the tropics (Lal, 2015),
43 and more than 3500 million hectare (Mha) of global land area (Karlen and Rice, 2015) are currently affected by
44 soil degradation, with serious implications for food security and the likelihood of malnutrition, ethnic conflict,
45 and civil unrest (Lal, 2009). In response to these problems, an increasing interest in soil degradation has been
46 observed among researchers and policy makers (Scherr 1999; Adesodun et al. 2008; Baumhardt et al. 2015;
47 Hueso-González et al. 2014; Lal, 2015; Teshahunegn, 2016).

48 Soil degradation is a measurable loss or reduction of the current or potential capability of soils to produce
49 plant materials of desired quantity and quality (Chen et al. 2002). Many scientists viewed soil degradation as a
50 decline in soil quality (Lal 2001; Adesodun et al. 2008; Beniston et al. 2015), and soil quality (SQ) as the capacity
51 of a soil to function within ecosystem and land-use boundaries (Doran and Zeiss, 2000; Karlen et al. 2001; Doran,
52 2002; Yemefack, 2005). Unfortunately, when soil degradation reaches an advance stage, soil quality restoration
53 is practically difficult (Lal and Cummings 1979). Therefore, good knowledge of SQ is important for developing
54 appropriate anti-degradation measures (Teshahunegn, et al., 2011). Since, soil degradation and soil quality are
55 interlinked through many processes (Lal, 2015), scholars have suggested that soil degradation can be assessed
56 using soil quality assessment strategies (Teshahunegn, 2014, Pulido et al. 2017). But, an essential step when
57 assessing soil degradation based on soil quality assessment strategies is the need for careful selection of
58 appropriate indicators relevant to degradation processes under investigation.

59 Degradation of soils is complex, often the consequence of many interacting processes (Prager et al.
60 2011). However, major processes include accelerated erosion (Cerde et al. 2009; Bravo-Espinosa et al. 2014);
61 deforestation (De la paix et al. 2013); poor pasture management (De Souza Braz et al. 2013); decline in soil
62 structure (Cerde 2000); salinization associated with inadequate irrigation management (Prager et al. 2011;
63 Ganjegunte et al. 2014); alkalization and sodification (Condom et al. 1999); depletion of soil organic matter
64 (SOM) (Novara et al. 2011); reduction in the activity of soil microorganisms (Lal 2009); and soil compaction
65 (Pulido et al. 2017). For sustainable soil management in agricultural regions, it is essential for farmers and
66 scientists to identify major dominant degradation processes and their indicators.

67 Cocoa (*Theobroma cacao* L.) is a major agricultural landuse type in the tropical rainforest belt of West
68 Africa (Tondoh et al. 2015), covering an estimated total area of about 6 million-ha in Côte d'Ivoire, Ghana,
69 Nigeria and Cameroon (Sonwa et al. 2004). Unfortunately, cocoa landscapes are often associated with a range of
70 ecological changes including deforestation, biodiversity loss, destruction of soil flora and fauna from pesticide
71 usage, and accelerated soil degradation (Critchley and Bruijnzeel 1996; Salami 1998, 2001; Rice and Greenberg
72 2000; Asare 2005; Ntiamoah and Afrane 2008; Mbile et al. 2009; Adeoye and Ayeni 2011; Jagoret et al. 2012;



73 Akinyemi 2013; Schoneveld 2014; Sonwa et al. 2014, Tondoh et al. 2015). Till date, soil degradation assessments
74 at plot scale in regions undergoing farmland conversion to cocoa agroforests are limited.

75 Worldwide, agricultural practices have been regarded as one of the major causes of soil degradation
76 (Kessler and Stroosnijder 2006, Rahmanipour, et al. 2014, Karlen and Rice, 2015, Zornoza et al., 2008) It is
77 widely acknowledged that agricultural practices or land use changes in agricultural regions alter key soil
78 properties such as soil organic matter (SOM), total nitrogen (TN), cation exchange capacity (CEC), exchangeable
79 cations, water holding capacity (WHC), bulk density (BD), and total porosity (TP) (Lemenih et al. 2005; Awiti
80 et al. 2008; Trabaquini 2015; Dawoe et al. 2010, 2014; Ameyan & Ogidiolu 1989; Hadgu et al. 2009; Thomaz &
81 Luiz 2012; Zhao et al. 2014; Tesfahunegn 2014). Although, many of these soil properties are regularly used as
82 indicators of soil degradation (Trabaquini 2015), the use of individual soil characteristics often provides an
83 incomplete representation of soil degradation (De la Rosa 2005; Puglisi et al. 2005, 2006). To overcome this
84 shortcoming, an integration of soil properties into numeric indices has been proposed (Doran & Parkin, 1994,
85 Leirós, et al. 1999; Bastida et al. 2006, Gómez et al. 2009, Puglisi et al. 2005, 2006; Sharma et al. 2008; Pulido
86 et al. 2017). Thus, Sánchez-Navarro et al. (2015) developed an overall soil quality index suitable for monitoring
87 soil degradation in semiarid Mediterranean ecosystems. Pulido et al. (2017) developed a soil degradation index
88 for rangelands of Extremadura SW Spain based on six indicators, cation exchange capacity (CEC), available
89 potassium, soil organic matter (SOM), water content at field capacity, soil depth and the thickness of the Ah-
90 horizon. Gomez et al. (2009) developed three soil degradation indexes obtained through a principal component
91 analysis (PCA) of the soils under organic olive farms in southern Spain. One of the index used only three soil
92 properties, organic C, water stable macroaggregates, and extractable P. According to these authors, this index has
93 the highest potential to be used as a relatively easy and inexpensive screening test of soil degradation in organic
94 olive farms in southern Spain. Till date, less attention has been given to development of numeric indices for
95 monitoring soil degradation under crop-specific landuse management systems in tropical countries. Whereas,
96 such indices can serve as the basis for integrating and interpreting several soil measurements, thereby indicating
97 whether a landuse management system is sustainable or not.

98 The aim of the present study is to develop a composite soil degradation assessment index (CSDI) for
99 shaded cocoa agroforests under tropical conditions in southwest Nigeria. This area is currently suffering from soil
100 degradation arising from cocoa based agroforests under a “slash and burn” farming system. Soil conditions under
101 age-sequenced peasant cocoa agroforests are investigated. The agroforest ages of 1-10 years, 11-40 years and 41-
102 80 years – hereafter referred to as young cocoa plantation (YCP), mature cocoa plantation (MCP) and senescent
103 cocoa plantation (SCP) respectively – were targeted as this is in line with the biological cycle of the cocoa tree
104 (Isaac et al. 2005; Jagoret et al. 2011, 2012; Saj et al. 2013). The specific objectives are: (i) to identify the most
105 important soil degradation processes associated with shaded cocoa agroforestry in the study area; (ii) to select a
106 minimum data set (MDS) of soil degradation indicators using multivariate statistical techniques; (iii) to integrate
107 the MDS into a CSDI; and (iv) to statistically validate CSDI and evaluate to what extent the CSDI can be used as
108 a tool by researchers, farmers, agricultural extension officers and government agencies involved in rehabilitation
109 of degraded cocoa soils in southwest Nigeria (and similar environments).



125 2.0 Materials and Methods

126 2.1 Study area

127 This study was carried out in the Ife region, southwest Nigeria (Figure 1), where most of the soils have been under
128 cocoa plantations for more than eighty years (Abiodun, 1971; Berry, 1974). The climate is humid tropical with a
129 mean daily minimum temperature of 25°C and a mean maximum temperature of 33°C. The mean annual rainfall
130 ranges between 1400 mm and 1600 mm, with a long- wet season lasting from April to October, and a relatively
131 short dry season that lasts from November to March. The natural vegetation is dominated by humid tropical
132 rainforests of the moist evergreen type, characterized by multiple canopies and lianas. The area is underlain by
133 rocks from the Basement complex of Pre-Cambrian Age, which are exposed as outcrops in several areas. The
134 soils are mainly Alfisols, classified as Kanhaplic Rhodustalf in the USDA Soil Taxonomy (Soil Survey Staff,
135 2006), or Luvisols (World Soil Reference, 2006) and locally known as Egbeda Association (Smyth &
136 Montgomery 1962). The area of study lies within the Egbeda soil series, characterised by sandy loam soils, with
137 increasing clay content in the lower horizons. The soils are slightly acidic to neutral in reaction (pH 6.5). With
138 the exception of the areas set aside as forest reserves, the natural vegetation has been replaced with perennial and
139 annual crops. Cocoa agroforests in the region were traditionally established using “slash and burn” approach
140 (Tondoh et al. 2015; Ngo-mbogba et al. 2015), where primary or secondary forests are selectively cleared, burned
141 and cocoa is planted along with understory food crops (Isaac et al. 2005). Farmers have recently shifted towards
142 full-sun cocoa agroforestry, particularly in areas where natural forest is scarce (Oke and Chokor 2009). Cocoa
143 trees are regularly sprayed with chemicals to combat black pod disease (*Phytophthora* sp), but farmers depend
144 entirely on the natural fertility of the soil without application of inorganic fertilizers or organic manure.

145 2.2 Site selection

146 A reconnaissance survey of Ife region was carried out between March and April 2013. Considering soil
147 variability and heterogeneity, five settlements of cocoa farmers (Mefoworade, Omifunfun, Aye Coker, Aba
148 Oyinbo and Kajola-Onikanga) in the southern Ife area were randomly selected as study sites. In each site, a total
149 of eight (8) cocoa stands of different ages (since site clearance) were randomly selected and assigned to three
150 cocoa plantation age categories: YCP (10 plots), MCP (15 plots) and SCP (15 plots). All sampled plots were
151 restricted to upper slope positions of a catena where the slope angle did not exceed 2° to ensure that catenary
152 variation in soil properties between the farms studied was minimal. Local farmers served as the main source of
153 information on the age distribution of the cocoa plantations and their permission was also sought to use their
154 farms as research plots. Each research plot was visited several times and notes were made on the physical
155 characteristics of the fields, their approximate sizes, presence of other crops and neighbouring trees, levels of
156 farm maintenance and evidence of soil erosion.

157

158 2.3 Soil sample collection for laboratory analysis

159 Soil sampling was conducted in May 2013. A quadrant measuring 1000 m² was demarcated at the centre of each
160 cocoa plantation. Each quadrant was subdivided into ten 100 m² sub-quadrants and serially labelled. Soil samples
161 were drawn at the centre of the even-numbered sub-quadrants, resulting in a total of five soil samples per plot.



162 Measurements were confined to the top 0–20 cm soils for the following reasons: (i) most significant changes in
163 soil characteristics in any vegetation (especially in a tropical environment) are confined to the topmost layer of
164 the soil profile (Aweto 1981; Aweto and Iyanda 2003); (ii) these depths cover the main distribution of roots and
165 soil nutrient stocks of cocoa plantations (Hartemink 2005); (iii) biological processes, such as earthworm activities
166 are restricted to 0–10 cm layer of tropical soils; (iv) to facilitate future replication of the methodology as routine
167 soil samples are usually kept at top-soil layer (plough layer). Two categories of soil samples were taken at each
168 sampling point to promote a detailed investigation of soil-property differences. The first was an undisturbed
169 sample using a bulk-density ring measuring 5 x 5 cm (diameter and height), whereas the other sample was taken
170 using a soil auger. The first sample was used to determine bulk density (BD), water-holding capacity (WHC) and
171 saturated hydraulic conductivity (SHC), and the second sample was used to determine the other studied soil
172 properties. The soil samples were stored in labelled polythene bags and taken to the laboratory for analysis. The
173 composite soil samples aggregated from the five samples collected in each plot were air-dried for two weeks,
174 hand ground in a ceramic mortar, passed through a 2 mm sieve and analysed for chemical properties and particle-
175 size distribution. Twenty-two soil properties were selected for analysis. The analytical methods are summarized
176 in Table 1.

177 **2.4 Statistical analyses and index development**

178 Based on extensive review of literature on soil quality and degradation assessment indexing, CSDI was developed
179 using a range of statistical techniques and procedures. The methodology consisted of eight steps as shown
180 schematically in fig. 2. Each of these steps is outlined below.

181 Step 1) involved selection of relevant indicators of soil degradation. Here, we selected twenty-two (22) analytical
182 soil properties widely acknowledged as soil quality and degradation indicators.

183 In Step 2) a factor analysis was performed to group all the soil data into statistical factors with principle component
184 analysis (PCA) as the method of factor extraction (Tesfahunegn et al., 2011). Factors were subjected to varimax
185 rotation with Kaiser normalization in order to generate factor patterns that load highly significant variables into
186 one factor, thereby producing a matrix with a simple structure that is easy to interpret (Ameyan and Ogidiolu
187 1989; de Lima et al. 2008; Momtaz et al. 2009). Factors with eigenvalues of less than one (1) were ignored. The
188 order in which the factors were interpreted was determined by the magnitude of their eigenvalues. Under each
189 factor, soil properties regarded as highly important were retained. These were defined as those that had a loading
190 value within 10% of the highest loading within an individual factor (Andrews et al. 2002). Soil properties that are
191 widely acknowledged as good indicators of soil quality, but with factor loading scores ≤ 0.70 , were also retained.
192 Soil physical, chemical and biological properties that have been suggested as important soil quality indicators
193 include soil organic carbon, available nutrients and particle size, bulk density, pH, soil aggregate stability, cation
194 exchange capacity and available water content (Doran and Parkin, 1994; Larson and Pierce, 1994; Karlen et al.,
195 1997; Zornoza et al., 2007; García-Ruiz et al., 2008; Qi et al., 2009; Marzaioli et al., 2010; Fernandes et al., 2011;
196 Lima et al., 2013; Merrill et al., 2013; Rousseau et al., 2013; Singh et al., 2014; Zornoza et al. 2015). In cases
197 where more than one soil property was found to be of high importance under a single PC, Pearson's correlation
198 coefficients were used to determine if any of these variables are redundant (Qi et al. 2009). When two highly



199 important variables were found to be strongly correlated ($r^2 > \pm 0.70$; $p < 0.05$), the one with the highest factor
 200 loading (absolute value) was retained (Andrews & Carroll 2001; Andrews et al. 2002; Montecchia et al. 2011).

201 In Step 3) of the CSDI development, the highly important soil properties under each factor were subjected to
 202 stepwise discriminant analysis (STEPDA) to select key soil properties (variables). In principle, stepwise
 203 discriminant analysis generates two or more linear combinations of the discriminating variables, often referred to
 204 as discriminant functions (Teschfahunegn et al., 2011). Whereas, the discriminant functions can be represented as:

$$205 \quad D_i = d_{i1}Z_1 + d_{i2}Z_2 + \dots + d_{ip}Z_p. \quad (\text{eq 1})$$

206 where D_i is the score on discriminant function i , the d 's are weighting coefficients, and the Z 's are the standardized
 207 values of the p discriminating variables used in the analysis (Awiti et al. 2008). In this study, STEPDA was used
 208 to select variables with the highest power to discriminate between the treatments. The validity of the result was
 209 evaluated using the Wilk's Lambda value. This value is an index of the discriminating power ranging between 0
 210 and 1 (the lower the value, the higher the discriminating power). At each step of STEPDA, the variable that
 211 minimizes the overall Wilks' Lambda was selected. One of the advantages of STEPDA is that the final model
 212 contains the variables that are considered useful. The result of this process was an MDS consisting of the most
 213 important variables for quantifying soil degradation in the selected plantations.

214 Step 4) involved the normalisation of the MDS variables to numerical scores between 0 and 1 using a linear
 215 scoring function (Masto et al. 2008; Ngo-mbogba et al. 2015). The "more is better" scoring curve was used to
 216 determine the linear score of soil variables:

$$217 \quad S_L = \left(\frac{x-l}{h-l} \right) \quad (\text{eq 2})$$

218 where, S_L is the linear score (between 0 and 1) of a soil variable, x is the soil variable value, l is the minimum
 219 value and h is the maximum value of soil variable.

220 During Step 5), the normalized MDS values were transformed into degradation scores (D) as described by Gómez
 221 et al. (2009) and obtained from:

$$222 \quad D = 1 - SL \quad (\text{eq 3})$$

223 where D is the degradation score and SL is the normalized MDS value. Here, a score of 1 signifies the highest
 224 possible soil degradation score and 0 represents complete absence of degradation for a particular soil property.

225 In Step 6) the degradation scores (D) were integrated into an index using the weighted additive method:

$$227 \quad \text{CSDI} = \sum_{i=1}^n (W_i D_i) \quad (\text{eq 4})$$

228 where CSDI represents the composite soil degradation index, W_i is the weight of variable i , D_i represents the
 229 degradation scores of the parameters in the MDS for each of the cocoa farms, and n is the number of indicators
 230 in the MDS. W_i in eq. [4] was derived by the percentage of the total variance explained by the factor in which
 231 the soil property had the highest load divided by the total variance explained by all the factors with eigenvalues
 232 ≥ 1 (Masto et al. 2008; Armenise et al. 2013).

233 In Step 7) CSDI values were categorized into number of desired (3) classes of degradation using their Z-score
 234 value as obtained by:



$$z = \frac{x - \mu}{\sigma} \quad (\text{eq 5})$$

235
236 where, Z is the z-score, x is the CSDI value of each plot, μ is the mean value and σ is the standard deviation. In
237 principle, z-scores explain the standard deviations of input values from the mean (Hinton 1999). For this purpose,
238 a Z values between -1 and 1 were regarded as having a *moderate* degradation status, while values of more than 1
239 was regarded as *high* and less than -1 as *low* (see results section for further explanation on this categorization).
240 In Step 8) the CSDI classification was statistically validated using a canonical discriminant analysis (CANDA).
241 Canonical discriminant analysis is a multivariate statistical technique whose objective is to discriminate among
242 pre-specified groups of sampling entities. The technique involves deriving linear combinations of two or more
243 discriminating variables (canonical variates) that will best discriminate among the *a priori* defined groups. In this
244 study, we used the “leave-one-out” cross validation procedure of CANDA. Using this procedure, a given
245 observation is deleted (excluded) and the remaining observations are used to compute a canonical discriminant
246 function that is used to assign the observation into a degradation class with the highest probability. For instance,
247 a sample with a probability of 0.003, 0.993 and 0.004 belonging to low, moderate and high degradation class
248 respectively was assigned to medium. This procedure is repeated for all observations and the result is a “hit ratio”
249 or confusion matrix, which indicates the proportions of observations that are correctly classified. Additionally,
250 CANDA was used to confirm the significance of the explanatory variables that discriminate between the three
251 soil degradation classes. In this study, the threshold (T) for the selection of variables correlating significantly with
252 the canonical discriminant functions was taken as $T = 0.2/\sqrt{\lambda}$ (eigenvalue) as suggested by Hadgu et al. (2009).
253 Scoring and indexing were performed using Microsoft Excel 2013. All statistical analyses were performed using
254 XLSTAT version 2016 (Addinsoft New York, USA).

255

256 3.0 Results and discussion

257 3.1 Identification of soil degradation processes using factor analysis

258 Table 2 shows the results of the factor analysis and reveals that the first five PCs had eigenvalues > 1 as illustrated
259 by the scree test (figure 3). Each PC explained 5% or more of the variation of the dataset. The first five PCs jointly
260 accounted for more than 77% of the total variance in the data set. In addition, it explained 68% of the variance in
261 available phosphorus, 84% in SOM, 76% in calcium, 65% in pH, 87% in clay, 90% in total nitrogen, 77% in silt,
262 83% in magnesium, 83% in sand, and 58% in bulk density. The high communalities among the soil properties
263 suggests that variability in selected soil properties is well accounted for by the extracted factors (Tesfahunegn *et*
264 *al.*, 2011).

265 Extractable zinc, extractable manganese and silt had high positive loadings on PC1 (0.875, 0.857, and
266 0.838 respectively). Because a significant correlation exists between extractable zinc and extractable manganese
267 ($r=0.834$, $p<0.001$; Table 3), the latter variable was excluded. For ease of association, PC1 was labelled *soil*
268 *micronutrient degradation factor*. PC2 was loaded highly by CEC (0.884) and exchangeable calcium (0.871), but
269 given that the correlation analysis showed a strong relationship ($r=0.870$, $p<0.001$; Table 3) between CEC and
270 exchangeable calcium, the latter was also excluded. SOM, with a relatively high factor loading (0.711), was
271 retained owing to its relevance in monitoring soil quality degradation (Brejda et al. 2000; Sharma et al.2009;
272 Masto et al. 2008; 2009; Zornoza, et al., 2015). Because the correlation coefficient between SOM and CEC was



273 relatively low ($r=0.578$; $p<0.001$; Table 3), both were retained as highly important variables. Given that SOM
274 was significantly correlated with several of the eliminated soil properties in the group, the second component
275 factor was labelled the *soil organic matter degradation factor*.

276 The third component factor (PC3) was highly loaded on available phosphorus (0.810) and total porosity
277 (0.801). Because the correlation coefficient between the two variables is relatively low ($r=0.578$; $p<0.001$; Table
278 3), both properties were retained. The group of variables associated with the third factor was termed the *available*
279 *phosphorus degradation factor*. The fourth factor was labelled as *soil acidity degradation factor* because it was
280 highly loaded on pH (0.791) only. Similarly, the fifth factor was labelled *soil textural degradation factor* because
281 it was dominated by clay (0.812).

282 So far, the PCA result suggests that soil degradation in the study region is mainly linked to four
283 degradation processes, namely 1) decline in soil nutrient, 2) loss of soil organic matter, 3) increase in soil acidity
284 and 4) the breakdown of soil textural characteristics arising from differences in clay eluviation (Figure 4). Figure
285 5 summarises the results of the interrelationship among the 22 soil properties as a correlation circle. The figure
286 shows that the first two PCA axes jointly accounted for 40.08 % of the total variance, with the first axis
287 (eigenvalue = 8.545) representing mainly micronutrients with extractable manganese, zinc, silt and total nitrogen
288 in contrast to bulk density, copper and sand. The second axis (eigenvalue = 3.96) is represented by CEC and
289 exchangeable calcium as opposed to the pH content of the soils. Figure 6 represents the percentage contributions
290 of the investigated soil properties in selected cocoa plantation chronosequence (CPC).

291

292 3.2 Selecting a minimum dataset (MDS) of soil degradation indicators

293 The PCA results presented thus far suggest that eight indicators (extractable zinc, silt, SOM, CEC,
294 available phosphorus, total porosity, pH, and clay) can be used to assess soil degradation in the study area.
295 However, the collection and analysis of such a large number of indicators is not viable for monitoring programmes
296 covering extensive areas and the identification of key soil degradation indicators will be very useful. The eight
297 soil properties were consequently subjected to forward stepwise discriminant analysis (STEPDA) to determine
298 which of them are most important for soil degradation monitoring in the study area. Figure 7 and Table 4 show
299 that STEPDA separated cocoa plantation chronosequence (CPC) into three groups (YCP, MCP and SCP), based
300 on the explanatory variables (8 soil parameters) included in the model. The first discriminant function separates
301 the MCP from YCP and SCP, while the second discriminant function separates YCP from MCP and SCP. The
302 overall Wilks' lambda test ($\lambda=0.047$; $p<0.001$) confirms that the means of the cocoa plantation
303 chronosequence (CPC) were significantly different for the two discriminant functions.

304 Table 4 shows that the first discriminant function which accounts for more than 80% of the variance in
305 soil properties is positively correlated with organic matter (0.952; $p<0.001$), extractable zinc (0.806; $p<0.001$),
306 CEC (0.611; $p<0.001$), thus it is labelled *soil organic matter and macro nutrients* dimension. This result suggests
307 that the plots in MCP have higher concentrations of soil nutrients than YCP and SCP. Similarly, the second
308 discriminant function, which accounts for more than 19% of the variance in soil properties is positively correlated
309 with CEC (0.622; $p<0.001$) and SOM (0.096), but negatively correlated with silt (0.520), clay (0.139), porosity



310 (0.309), zinc (0.527), and available phosphorus (0.035). This suggests that the YCP cases have poor physical soil
 311 properties compared to MCP and SCP. This function is labelled *soil physical and micronutrient dimension*.

312 The result of STEPDA confirmed that only four soil properties are significant in discriminating between
 313 the cocoa plantation chronosequence (CPC). These soil properties and their partial regression (R^2) are SOM
 314 ($R^2=0.797$, $p<0.001$; Wilks' Lambda=0.203), extractable zinc ($R^2=0.548$, $p<0.001$; Wilks' Lambda=0.259), CEC
 315 ($R^2=0.379$, $p<0.001$; Wilks' Lambda=0.432) and clay ($R^2=0.169$, $p<0.05$; Wilks' Lambda=0.866). The relative
 316 importance of these variables, as indicated by the length of their eigenvectors, is (in decreasing order) SOM,
 317 extractable zinc, CEC, and clay. Consequently, these four soil properties constitute a minimum dataset (MDS) of
 318 soil degradation indicators in our study area.

319 3.3 MDS normalization, transformation and integration into CSDI

320 The four selected indicators of the MDS were normalized and transformed into degradation scores (D) as
 321 described in Section 2.4. Weights were assigned to each degradation score using the result of the factor analysis
 322 (Table 2). As an example, the procedure to calculate the weighting factor for extractable zinc was as follows: the
 323 individual percentage variance for PC1 (23.70), was divided by 77.15%, the cumulative percentage of variation
 324 explained by all the retained PCs (Table 3), to yield the weight of 0.31. After assigning different weights to each
 325 parameter, they were integrated into a CSDI. This index is the sum of the normalised and weighted values of each
 326 parameter. CSDI was computed for each cocoa agroforests as:

$$327 \text{CSDI} = 0.21 (\text{DSOM}) + 0.31 (\text{DZn}) + 0.21 (\text{DCEC}) + 0.17 (\text{DClay}) \quad (\text{eq 6})$$

328 Ordering the variables included in the equation as a function of the loading of the coefficient gave:

$$329 \text{CSDI} = 0.31 (\text{DZn}) + 0.21 (\text{DSOM}) + 0.21 (\text{DCEC}) + 0.17 (\text{DClay}) \quad (\text{eq 7})$$

330 where, CSDI is the composite soil degradation index and DZn, DSOM, DCEC and DClay are the degradation
 331 scores of extractable zinc, organic matter, CEC and clay respectively.

332 3.4 Classification into degradation classes

333 Table 5 shows the soil degradation classification of CSDI scores by solving equation 5. In our case, μ and σ were
 334 calculated as 0.289 and 0.094 respectively, resulting in CSDI values of 0.195 when $Z = -1$ and 0.383 when $Z = 1$.
 335 Consequently, the CSDI classes are *Low* (<0.195) and *High* (>0.383). CSDI values between 0.195 and 0.383
 336 were regarded as *Moderate*. The interpretations of these classes is shown in table 6 (modified from Gómez et al.
 337 2009). Most (65%) of the selected cocoa farms are moderately degraded, while 18% have a high degradation
 338 status (Table 5). A significant difference was observed in the degradation status of YCP, MCP and SCP (ANOVA
 339 test, $F_{2,39}=57.59$; $P<0.001$). Fig 8 shows that 30% of YCP, 53.33% of MCP, and 100% of SCP are moderately
 340 degraded. However, 70% of YCP is highly degraded and 47% of MCP show no sign of degradation. This implies
 341 that MCP plots are less degraded compared to YCP and SCP. This result is consistent with other studies in West
 342 Africa. For instance, Dawoe et al. (2014) reported that, in humid lowland Ghana, soil properties and quality
 343 parameters of a ferric lixisol improved under cocoa plantations that have been operating for 15-30 years and were
 344 better than that of young cocoa plantations with a three-year production age. Similar results were obtained by
 345 Tondoh et al. (2015), who reported that, in Côte d'Ivoire, there was a steady degradation of soil quality over time
 346 in full-sun cocoa stands planted on ferralsols for 10 years, but the degradation value was less pronounced in 20-



347 year-old plantations. Comparing our results with those of Dawoe et al. (2014) and Tondoh et al. (2015) highlights
348 the effects of poor and unsustainable land management practices on soil degradation in peasant cocoa agroforests
349 in West Africa. Traditionally, cocoa plots are cultivated with food crops in the first three to five years of
350 development until the canopies have formed. Given that smallholder cacao farmers in the study area do not use
351 chemical fertilizers to improve soil quality, degradation of the physical, chemical and biological properties of
352 cocoa soils are imminent during this phase of plantation establishment.

353 3.5 Statistical validation of CSDI

354 A canonical discriminant analysis (CANDA) was used to validate the CSDI classification. The values of
355 the four soil properties (organic matter, extractable zinc, CEC and clay) were used as data input. Fig. 9 and Table
356 7 show that the three soil degradation classes (*low*, *moderate* and *high*) were significantly separated on the first
357 and second canonical functions (Wilk's Lambda=0.156, $F_{6,68}=13.04$, $p<0.0001$). Of the total variance, 93.46%
358 was accounted for by the first canonical function, which was significant at $p<0.001$. The second canonical
359 function accounted for 6.54% of the total variance and was significant at $P<0.005$. Extractable zinc, organic matter
360 and cation exchange capacity significantly contributed to the distinction among soil degradation classes and were
361 positively associated with the first canonical function (Table 7). Clay also contributed significantly to the
362 distinction among soil degradation classes, but was positively associated with the second canonical function
363 (Table 7).

364 CANDA classification results in Table 8 reveals that the CSDI model performs reasonable well, showing a
365 low level of misclassification. The table shows that for the original grouped cases, the CANDA correctly classified
366 6 of the 7 (85.7%) low, 23 of 26 (88.4%) moderate and all of the high cases. The implication of the CANDA
367 accuracy assessment is that the proposed classes of soil degradation (*Low*, *Moderate* and *High*) were significantly
368 separated by the four canonical variables included in the model and that the model can consequently be used with
369 a high degree of confidence. Result from this study indicate that the CSDI can effectively be used to monitor and
370 evaluate the degree of soil (Alfisols) degradation under cocoa plantation in the study area (and similar
371 environments). However, more work is needed, to apply and evaluate the index on different soil types from
372 different cocoa producing regions or countries.

373 4.0 Conclusions

374 In this study, we developed a composite soil degradation index (CDSI) to cost-effectively assess the status
375 of soil degradation under cocoa agroforests. Of the initial twenty-two (22) soil properties evaluated, multivariate
376 statistical analyses revealed that four (4) soil properties (extractable zinc, SOM, CEC and clay) were the main
377 indicators of soil degradation. This minimum dataset (MDS) of soil degradation indicators was used to produce a
378 CSDI, which was classified into three classes of degradation. According to this classification 65% of the selected
379 cocoa farms are moderately degraded, 17.5% have a high degradation status and 17.5% show no sign of
380 degradation. This classification corresponded well with a CANDA classification performed on the same dataset.

381 The findings suggest that the selection of a small set of relevant indicators will be more cost-efficient and
382 less time consuming than using a large number of soil properties that may be irrelevant to the processes of



383 degradation. They also suggest that soil degradation under cocoa agroforests (in this region at least) is mainly
 384 attributed to a decline in soil nutrient, loss of soil organic matter, increase in soil acidity and the breakdown of
 385 soil textural characteristics over time. This study shows that both physical and chemical soil properties are
 386 degraded under long-term cocoa production. The implications are serious for cocoa production sustainability on
 387 acidic Alfisols. Degradation of physical components of these soils portends serious risks to crop yields.
 388 Degradation of chemical soil properties, coupled with non-application of fertilizers, will likely exacerbate soil
 389 degradation processes. To prevent smallholder cocoa production from becoming unsustainable in the long-term,
 390 it is critical to advise farmers of the need for the application of artificial fertilizers, particularly under young cocoa
 391 plantations. Although the application of fertilizers will substantially improve the soil structure and nutrient
 392 conditions of cocoa soils, the poor transportation system in rural areas and prohibitive costs associated with
 393 artificial fertilizer application in cocoa groves remains a challenge to both farmers and government.

394 **5.0 Acknowledgement**

395 Financial support provided by the TETFund, administrated by the Osun State University Research Committee, is
 396 gratefully acknowledged. A special word of gratitude is owing to Dr Kayode Are, soil physicist at the Institute of
 397 Agricultural Training, Obafemi Awolowo University, for his assistance during fieldwork. The efforts of the
 398 technical and laboratory staff of Soil and Land Resource Management, Obafemi Awolowo University, Ile-Ife,
 399 Nigeria are sincerely acknowledged. We are also grateful to the chiefs of the various villages for their support
 400 during the interviews and the forty cocoa farmers for their permission to carry out this study on their farms.

401

402

403 **6.0 References**

404 Abiodun, J.: Service centres and consumer behaviour within the Nigerian Cocoa Area. *Geografiska Annaler series*
 405 *B, Human Geography*, 53(2),78–93, 1971.

407 Adejuwon, J.O., and Jeje, L.K.: Land element of the environmental system of Ife area. Occasional Publication,
 408 Department of Geography, University of Ife, 1975.

409 Adejuwon, J.O., and Ekanade, O.: A comparison of soil properties under different landuse types in a part of the
 410 Nigerian cocoa belt, *Catena*, 15, 319–331, 1988.

411 Adeoye, N.O., and Ayeni, B.: Assessment of deforestation, biodiversity loss and the associated factors: case
 412 study of Ijesa-Ekiti region of Southwestern Nigeria, *GeoJournal* 76: 229–243. doi:10.1007/s10708-009-
 413 9336-z, 2011.

414 Adesodun, J.K., Davidson, D.A., and Mbagwu, J.S.C.: Soil quality assessment of an oil-contaminated tropical
 415 Alfisol amended with organic wastes using image analysis of pore space, *Geoderma*, 146, 166–74,
 416 doi:10.1016/j.geoderma.2008.05.013, 2008.

417 Akinyemi, F.O. : An assessment of landuse change in the cocoa belt of south-west Nigeria, *Int. J. Remote Sens.*,
 418 34, 2858–2875, 2013.



- 419 Ameyan, O., and Ogidiolu, O.: Agricultural landuse and soil degradation in a part of Kwara State, Nigeria,
420 Environmentalist, 9, 285–290, 1989.
- 421 Anderson, J.M., and Ingram, J.S.I. (eds): Tropical soil biology and fertility: a handbook of methods. CAB
422 international. Wallingford, UK, 1993 .
- 423 Andrews, S.S., and Carroll, C.R.: Designing a soil quality assessment tool for sustainable agroecosystem
424 management, Ecol. Appl., 11,1573–1585, 2001.
- 425 Andrews, S.S., Karlen, D.L., and Mitchell, J.P.: A comparison of soil quality indexing methods for vegetable
426 production systems in Northern California, Agr. Ecosyst Environ., 90, 25–45, doi:10.1016/S0167-
427 8809(01)00174-8, 2002.
- 428 Areola, O.: Extractable copper content of soils under peasant cocoa farms in Ibadan region, Nigeria, Turrialba,
429 35, 229–232, 1985.
- 430 Armenise, E., Redmile-Gordon, M.A., Stellacci, A.M., Ciccarese, A., and Rubino, P.: Developing a soil quality
431 index to compare soil fitness for agricultural use under different managements in the Mediterranean
432 environment, Soil Till. Res., 130, 91–98, doi:10.1016/j.still.2013.02.013, 2013.
- 433 Asare, R.: Cocoa agroforests in West Africa: a look at activities on preferred trees in the farming systems.
434 Forestry and Landscape Working Paper, Arboretum Working Paper, No. 6. Forest and Landscape Denmark,
435 2005.
- 436 Aweto, A.O.: Organic matter in fallow soil in a part of Nigeria and its effects on soil properties. J. Biogeogr., 8:
437 67–74, 1981.
- 438 Aweto, A.O., and Iyanda, A.O.: Effects of *Newbouldia Laevis* on soil subjected to shifting cultivation in the
439 Ibadan Area, Southwestern Nigeria, Land Degrad. Dev., 56, 51–56, 2003.
- 440 Awiti, A.O., Walsh, M.G., Shepherd, K.D., and Kinyamario, J.: Soil condition classification using infrared
441 spectroscopy: A proposition for assessment of soil condition along a tropical forest-cropland
442 chronosequence, Geoderma, 143, 73-84, 2008.
- 443 Bastida, F, Luis M.J., and García C.: Microbiological degradation index of soils in a semiarid climate, Soil Biol.
444 Biochem., 38: 3463-3473. doi:10.1016/j.soilbio.2006.06.001, 2006.
- 445 Baumhardt, R.L., Stewart, B.A., and Sainju, U.M.: North American soil degradation: processes, practices, and
446 mitigating strategies, Sustainability , 7: 2936-2960, 2015.
- 447 Beniston, J.W., Lal, R., and Mercer, K.L.: Assessing and managing soil quality for urban agriculture in a degraded
448 vacant lot soil, Land Degrad. Dev. doi:10.1002/ldr.2342, 2015.
- 449 Berry, S.: The concept of innovation and the history of cocoa farming in western Nigeria. The Journal of African
450 History. 15(1),83–95, 1974.
451
- 452 Bravo-Espinosa, M., Mendoza, M.E., Carlón-Allende, T., Medina, L., Sáenz-Reyes, J.T., and Páez, R.: Effects of
453 converting forest to avocado orchards on topsoil properties in the Trans-Mexican volcanic system, Mexico,
454 Land Degrad. Dev., 25, 452-467, 2014.



- 455 Bray, R.H., and Kurtz, L.T.: Determination of total organic and available forms of phosphorus in soils, *Soil Sci.*,
 456 59, 39-45, 1945.
- 457 Brejda, J. J., Karlen, D.L., Smith, J. L., and Allan, D.L.: Identification of regional soil quality factors and
 458 indicators: II. Northern Mississippi Loess Hills and Palouse Prairie, *Soil Sci Soc Am. J.*, 64, 2125–2135,
 459 2000.
- 460 Brevik, E.C., Cerdà A., Mataix-Solera, J., Pereg, L., Quinton, J.N., Six, J., and Van Oost, K.: The interdisciplinary
 461 nature of SOIL, *SOIL 1*: 117-129, doi:10.5194/soil-1-117-2015, 2015.
- 462 Cambardella, C.A., Gajda, A.M., Doran, J.W., Wienhold, B.J., and Kettler, T.A.: Estimation of particulate and
 463 total organic matter by weight loss-on-ignition. In: Lal, R., Kimbe, J.M., Follet, R.F., and Stewart, B.A.
 464 (eds), *Assessment methods for soil carbon*. Boca Raton (FL): Lewis Publishers. 349–359, 2001.
- 465 Condom, N., Kuper, M., Marlet, S., Valles, V., and Kijne, J.: Salinization, alkalinization and sodification in
 466 punjab (pakistan): characterization of the geochemical and physical processes of degradation, *Land Degrad.*
 467 *Dev.*, 10, 123-140, 1999.
- 468 Cerdà, A.: Aggregate stability against water forces under different climates on agriculture land and scrubland in
 469 southern Bolivia, *Soil Till. Res.*, 57: 159-166, 2000.
- 470 Cerdà, A., Morera A.G., and Bodi, M.B.: Soil and water losses from new citrus orchards growing on sloped soils
 471 in the western, *Earth surf. processes*, 34: 1822-1830, 2009.
- 472 Chen, J., Chen, J., Tan, M., Gong, Z.: Soil degradation : a global problem endangering sustainable development.
 473 *Journal of Geographical Sciences*, 12, 2: 243–252, 2002.
- 474 Chude, V.O.: The nutritional status of cacao (*theobroma cacao L.*) with respect to boron and zinc in soils of south-
 475 western Nigeria. PhD thesis, University of Ibadan, 1983.
- 476 Critchley, W., and Bruijnzeel, L.A.: Environmental impacts of converting moist tropical forest to agriculture and
 477 plantations. UNESCO International Hydrological Programme accessed at
 478 <http://unesdoc.unesco.org/images/0010/001096/109608eo.pdf>, 1996.
- 479 de Lima A.C.R., Hoogmoed W., and Brussaard, L.: Soil quality assessment in rice production systems:
 480 establishing a minimum data set, *J. Environ. Qual.*, 37, 623-630, doi:10.2134/jeq2006.0280, 2008.
- 481 Dawoe, E.K., Isaac, M.E., and Quashie-Sam, J.: Litterfall and litter nutrient dynamics under cocoa ecosystems in
 482 lowland humid Ghana, *Plant Soil*, 330: 55–64, 2010.
- 483 Dawoe, E. K., Quashie-Sam, J.S., and Oppong S.K.: Effect of landuse conversion from forest to cocoa agroforest
 484 on soil characteristics and quality of a Ferric Lixisol in lowland humid Ghana, *Agroforestr Syst*, 88, 87–
 485 99, doi:10.1007/s10457-013-9658-1, 2014.
- 486 De la paix, M. J., Lanhai, L., Xi, C., Ahmed, S., and Varenayam, A.: Soil degradation and altered flood risk as a
 487 consequence of deforestation, *Land Degrad. Dev.*, 24, 478–485, 2013.
- 488 De la Rosa, D.: Soil quality evaluation and monitoring based on land evaluation, *Land Degrad. Dev.*, 16, 551–
 489 559, 2005.



- 490 De Souza Braz A.M., Fernandes A.R., and Alleoni L.R.F.: Soil attributes after the conversion from forest to
491 pasture in Amazon, *Land Degrad. Dev.*, 24, 33-38, 2013.
- 492 Doran, J.W. and Parkin, T. B.: Defining and Assessing Soil Quality, in: Defining soil quality for a sustainable
493 environment, edited by: Doran, J.W., Coleman, D. F., Bezdicek, D. F., and Stewart, B. A., *Soil Sci. Soc.
494 Am.*, Special Publication 35, Madison, WI, 3–21, 1994.
- 495 Driesien, I.H.O.: Patterns of land holding and land distribution in Ife Region, *African* 41,42-53, 1971.
- 496 Ekanade, O.: The impact of cocoa cultivation on soil characteristics in southwestern Nigeria. unpublished PhD.
497 Thesis, Department of Geography, University of Ife, Ile-Ife, Nigeria, 1985.
- 498 Ekanade, O. Small-scale cocoa farmers and environmental change in the tropical rain forest regions south-western
499 Nigeria, *J. Environ. Manage.*, 25: 61–70. 1987.
- 500 Falade, J.A.: Soil bulk density moisture supply interaction in Amazon Cocoa, *West African Journal of Biological
501 and Applied Chemistry*, 18:15–22, 1975.
- 502 Faniran, A., and Areola, O. (eds): Essentials of soil study (with special reference to tropical areas). Heineman,
503 London, 1978.
- 504 Fernandes, J.C., Gamero, C.A., Rodrigues, J.G.L., and Mirás-Avalos, J.M., Determination of the quality index of
505 a Paleudult under sunflower culture and different management systems. *Soil Till. Res.* 112, 167–174, 2011.
- 506 Ganjegunte, G.K., Sheng Z., and Clark, J.A.: Soil salinity and sodicity appraisal by electromagnetic induction in
507 soils irrigated to grow cotton, *Land Degrad. Dev.*, 25: 228-235, doi: 10.1002/ldr.1162, 2014.
- 508 García-Ruiz, R., Ochoa, V., Hinojosa, M. B., and Carreira, J. A.: Suitability of enzyme activities for the
509 monitoring of soil quality improvement in organic agricultural systems, *Soil Biol. Biochem.*, 40, 2137–
510 2145, 2008.
- 511 Gee, G.W., and Or, D.: Particle-size analysis. In: methods of soil analysis, Part 4. soil physical properties,
512 agronomy monograph 5. Dane, J. H., and Topp, G. C. (eds.) SSSA, Madison, WI, 225-275, 2002.
- 513 Grossman, R.B., and Reinsch, T.G.: Bulk density and linear extensibility: core method. In: Dane, J.H., Topp, G.C.
514 (eds). Methods of soil analysis. Part 4. Physical methods. Madison (WI), Soil Science Society of America.
515 208–228, 2002.
- 516 Gómez, J.A., Sonia, Á., and María-Auxiliadora, S.: Development of a soil degradation assessment tool for organic
517 Olive groves in Southern Spain, *Catena*, 79, 9–17, 2009.
- 518 Hadgu, K.M., Rossing, W.A., Kooistra, L., and van Bruggen, A.H.: Spatial variation in biodiversity, soil
519 degradation and productivity in agricultural landscapes in the highlands of Tigray, Northern Ethiopia. *Food
520 Security* 1, 83–97, doi:10.1007/s12571-008-0008-5, 2009.
- 521 Hartemink, A.E. Nutrient stocks, nutrient cycling, and soil changes in cocoa ecosystems: A Review. *Adv. Agron.*,
522 86 : 227–253, 2005.



- 523 Hueso-González, P., Martínez-Murillo J.F., and Ruiz-Sinoga, J.D.: The impact of organic amendments on forest
524 soil properties under Mediterranean climatic conditions, *Land Degrad. Dev.*, doi: 10.1002/ldr.2296, 2014.
- 525 Hinton, P.R.: *Statistics explained: A guide for social science students*. NY: Routledge, 1999.
- 526 Isaac, M.E., Gordon, A.M., Thevathasan, N., Oppong, S.K., and Quashie-Sam, J.: Temporal changes in soil
527 carbon and nitrogen in West African multistrata agroforestry systems: a chronosequence of pools and fluxes,
528 *Agroforestry Systems*, 65, 23–31, 2005.
- 529 Jagoret, P., Michel-Dounias, I., and Malézieux, E.: Long-term dynamics of cocoa agroforests: a case study in
530 central Cameroon, *Agroforestry Systems* 81: 267–278. DOI:10.1007/s10457-010-9368-x, 2011.
- 531 Jagoret, P., Michel-Dounias, I., Snoeck, D., Ngnogué, H.T., and Malézieux, E.: Afforestation of savannah with
532 cocoa agroforestry systems: a small-farmer innovation in central Cameroon, *Agroforestry Systems*, 86,
533 493–504. doi:10.1007/s10457-012-9513-9, 2012.
- 534 Karlen, D.L., Mausbach, M.J., Doran, J.W., Cline, R.G., Harris, R.F., Schuman, G.E., Soil quality: a concept,
535 definition, and framework for evaluation. *Soil Sci. Soc. Am. J.* 61, 4–10, 1997.
- 536
537 Karlen, D.L. and Rice, C.W.: Soil degradation: Will humankind ever learn? *Sustainability* 7(9):12490–12501,
538 2015.
- 539
540 Keesstra, S.D., Geissen, V., Mosse, K., Piirainen, S., Scudiero, E., Leistra, M., and van Schaik, L.: Soil as a filter
541 for groundwater quality, *Current Opinions in Environmental Sustainability* 4, 507-516,2012
- 542
- 543 Kessler, C. A., and Stroosnijder, L.: Land degradation assessment by farmers in Bolivian mountain valleys, *Land*
544 *Degrad. Develop.* 17: 235–248 ,2006.
- 545 Lal, R.: Soil degradation by erosion, *Land Degrad. Dev.*, 12 : 519–39, 2001.
- 546 Lal, R.: Soil degradation as a reason for inadequate human nutrition. *Food Security*, 1:45–57, 2009.
- 547
548 Lal, R.: Restoring soil quality to mitigate soil degradation. *Sustainability*, 7(5):5875–5895, 2015.
- 549 Lal R, and Cummings, D.J.: Clearing a tropical forest I. Effects on soil and micro-climate, *Field Crops Research*
550 2: 91–107, 1979.
- 551 Larson, W. E. and Pierce, F. J.: The dynamics of soil quality as a measure of sustainable mangement, in: *Defining*
552 *soil quality for a sustainable environment*, edited by: Doran, J.W., Coleman, D. C., Bezdiecek, D. F., and
553 Stewart, B. A., SSSA-Special Publica- tion 35, Soil Science Society of America, Madison, WI, 37–51, 1994
- 554 Leirós, M.C., Trasar-Cepeda, C., García-Fernández, F. and Gil-Sotres, F.: Defining the validity of a biochemical
555 index of soil quality. *Biology and Fertility of Soils.* 30(1-2):140–146, 1999.
- 556 Lemenih, M., Karlun, E., and Olsson, M.: Soil organic matter dynamics after deforestation along a farm field
557 chronosequence in southern highlands of Ethiopia, *Agriculture, Ecosystems & Environment* 109: 9–19,
558 doi:10.1016/j.agee.2005.02.015, 2005.



- 559 Lima, A.C.R., Brussaard, L., Totola, M.R., Hoogmoed, W.B., and de Goede, R.G.M.: A functional evaluation of
560 three indicator sets for assessing soil quality. *Appl. Soil Ecol.* 64, 194–200, 2013.
- 561 Marzaioli, R., D’Ascoli, R., De Pascale, R. A., and Rutigliano, F. A.: Soil quality in a Mediterranean area of
562 Southern Italy as re- lated to different land use types, *Appl. Soil Ecol.*, 44, 205–212, 2010
- 563 Masto, R.E., Chhonkar, P.K., Singh, D., and Patra, A.K.: Alternative soil quality indices for evaluating the effect
564 of intensive cropping, fertilisation and manuring for 31 years in the semi-arid soils of India. *Environmental
565 Monitoring and Assessment* 136: 419–435, 2008.
- 566 Masto, R.E., Chhonkar P.K., Singh, D., and Patra, A.K.: Changes in soil quality indicators under long-term
567 sewage irrigation in a sub-tropical environment, *Environmental Geology*, 56, 1237–1243,
568 doi:10.1007/s00254-008-1223-2, 2009.
- 569 Mbile, P., Ngaunkam, P., Besingi, M., Nfoumou, C., Degrande, A., Tsobeng, A., Sado, T., and Menimo, T.:
570 Farmer management of cocoa agroforests in Cameroon: Impacts of decision scenarios on structure and
571 biodiversity of indigenous tree species, *Biodiversity*, 10, 4, 12–19, doi:10.1080/14888386.2009.9712857,
572 2009.
- 573 Merrill, S.D., Liebig, M.A., Tanaka, D.L., Krupinsky, J.M., and Hanson, J.D.: Comparison of soil quality and
574 productivity at two sites differing in profile structure and topsoil properties. *Agriculture. Ecosyst. Environ.*
575 179, 53–61, 2013.
- 576 Milgroom, J., Gomez, J.A., Soriano, M.A., and Fereres, E.: From experimental research to an on-farm tool for
577 participatory monitoring and evaluation : an assessment of soil erosion risk in organic olive orchards, *Land
578 Degrad. Dev.*, 18, 397–411, 2007.
- 579 Momtaz, H.R., Jafarzadeh, A.A., Torabi, H., Oustan, S., Samadi, A., Davatgar, N., and Gilkes R.J.: An
580 assessment of the variation in soil properties within and between landform in the Amol region, Iran,
581 *Geoderma* 149: 10–18, 2009.
- 582 Montecchia, M.S., Correa, O.S., Soria, M.A., Frey, S.D., Garcia, A.F., and Garland, J.L.: Multivariate approach
583 to characterizing soil microbial communities in pristine and agricultural sites in Northwest Argentina,
584 *Applied Soil Ecology*, 47, 176–183, doi:10.1016/j.apsoil.2010.12.008, 2011.
- 585 Ngo-mbogba, M., Yemefack, M., and Nyeck, B.: Assessing soil quality under different land cover types within
586 shifting agriculture in South Cameroon, *Soil and Tillage Research*, 150,124–131, 2015.
- 587 Novara, A., Gristina, L., Bodi, M.B., and Cerdà A. The impact of fire on redistribution of soil organic matter on
588 a Mediterranean hillslope under maquia vegetation type, *Land Degrad. Dev.*, 22, 530-536. doi:
589 10.1002/ldr.1027, 2011.
- 590
591 Ntiamoah, A. and Afrane, G.: Environmental impacts of cocoa production and processing in Ghana: life cycle
592 assessment approach, *Journal of Cleaner Production* 16 1735-1740, 2008.
- 593
594 Oke, O. C., and Chokor, J. U.: Land snail populations in shade and full-sun cocoa plantations in South Western
595 Nigeria, West Africa, *African Scientist*, 10, 1, 19-29, 2009.
- 596
597 Olsen, S.R., and Sommers, L.E.: Phosphorus. In Sparks, D.L., Page, A.L., Helmke, P.A., and Loeppert, R.H. (eds)
598 method of soil analysis: chemical and microbiological properties, Part 2, agronomy monograph 9, 403 -
599 430. Soil Science Society of America, Wisconsin, WI, 1982



- 600 Peech, M. Hydrogen-ion activity. In methods of soil analysis. Black, C.A. (ed), 2, 914-926, 1965.
- 601 Prager, K., Schuler, J., Helming, K., Zander, P., Ratering, T., and Hagedorn, K.: Soil degradation, farming
 602 practices, institutions and policy responses: an analytical framework, *Land Degrad. Dev.*, 22, 32–46, 2011.
- 603 Puglisi, E., Nicelli, M., Capri, E., Trevisan, M., and Del Re, A.A.M.: A soil alteration index based on
 604 phospholipid fatty acids, *Chemosphere*, 61, 1548-1557, 2005.
- 605 Puglisi, E., Del Re, A.A.M., Rao, M.A., and Gianfreda, L.: Development and validation of numerical indexes
 606 integrating enzyme activities of soils, *Soil Biology and Biochemistry*, 38, 1673-1681,
 607 doi:10.1016/j.soilbio.2005.11.021, 2006.
- 608 Pulido, M., Schnabel, S., Contador, J.F.L., Lozano-Parra, J. and Gómez-Gutiérrez, Á.: Selecting indicators for
 609 assessing soil quality and degradation in rangelands of Extremadura (SW Spain). *Ecological Indicators*. 74,
 610 49–61, 2017.
- 611 Qi, Y., Darilek, J.L., Huang, B., Zhao, Y., Sun, W., and Gu, Z.: Evaluating soil quality indices in an agricultural
 612 region of Jiangsu Province, China, *Geoderma*, 149, 325-334, doi:10.1016/j.geoderma.2008.12.015, 2009.
- 613 Rahmanipour, F., Marzaioli, R., Bahrami, H.A., Fereidouni, Z. and Bandarabadi, S.R. Assessment of soil quality
 614 indices in agricultural lands of Qazvin Province, Iran. *Ecological Indicators*. 40, 19–26, 2014.
- 615 Reynolds, W.D., and Elrick, D.: Constant head soil core (tank) method. In: Dane, J.H., and Topp, G.C., (eds).
 616 Methods of soil analysis. Part 4. Physical methods. Madison (WI): Soil Science Society of America, 804–
 617 808, 2002.
- 618 Rice, R.A., and Greenberg, R.: Cacao cultivation and the conservation of biological diversity, *Ambio: A Journal*
 619 *of the Human Environment*, 29, 3, 20–25, 2000
- 620 Rousseau, L., Fonte, S.J., Téllez, O., van der Hoek, R., and Lavelle, P.: Soil macrofauna as indicators of soil
 621 quality and land use impacts in smallholder agroecosystems of western Nicaragua. *Ecological Indicators*.
 622 27, 71–82, 2013..
- 623 Saj, S., Jagoret, P., and Ngogue, H.T.: Carbon storage and density dynamics of associated trees in three contrasting
 624 Theobroma cacao agroforests of Central Cameroon, *Agroforestry Systems*, 87, 1309–1320,
 625 doi:10.1007/s10457-013-9639-4, 2013.
- 626 Salami, A.T.: Vegetation modification and man-induced environmental change in rural southwestern Nigeria.
 627 *Agriculture, Ecosystems and Environment* 70: 159–167, 1998.
- 628 Salami, A.T.: Agricultural colonisation and floristic degradation in Nigeria's rainforest ecosystem.
 629 *Environmentalist* 21 : 221–229, 2001.
- 630 Sánchez-Navarro, A., Gil-Vázquez, J.M., Delgado-Iniesta, M.J., Marín-Sanleandro, P., Blanco-Bernardeau, A.,
 631 and Ortiz-Silla, R.: Establishing an index and identification of limiting parameters for characterizing soil
 632 quality in Mediterranean ecosystems. *Catena* 131, 35–45, 2015
- 633 Scherr, S.J.: Soil degradation: a threat to developing country food security by 2020? vision 2020: food, agriculture,
 634 and the environment discussion paper 27, 14-25, 1999.



- 635 Schoneveld, G.C.: The politics of the forest frontier: Negotiating between conservation, development, and
636 indigenous rights in Cross River State, Nigeria, *Land Use Policy* 38, 147–162,
637 doi:10.1016/j.landusepol.2013.11.003, 2014.
- 638 Schulte, R.P., Bampa, F., Bardy, M., Coyle, C., Fealy, R., Gardi, C., Ghaley, B.B., Jordan, P., Laudon, H.,
639 O'Donoghue, C., and Ó'hUallacháin, D.: Making the most of our land: managing soil functions from local
640 to continental scale, *Frontiers in Environmental Science*, 3, 1-14, 2015.
- 641 Sharma, K.L., Mandal, U.K., Srinivas, K., Vittal, K.P., Mandal, B., Grace, J.K., and Ramesh, V.: Long-term soil
642 management effects on crop yields and soil quality in a dryland Alfisol, *Soil and Tillage Research*, 83, 246–
643 259, 2005.
- 644 Sharma, K.L., Grace, J.K., Mandal, U.K., Gajbhiye, P.N., Srinivas, K., Korwar, G.R., Hima Bindu, V., Ramesh,
645 V., Ramachandran, K., and Yadav, S. K.: Evaluation of long-term soil management practices using key
646 indicators and soil quality indices in a semi-arid tropical Alfisol, *Soil Research*, 46,368–37, 2008.
647
- 648 Sharma, K.L., Raju, K.R., Das, S.K., Rao, B.P., Kulkarni, B.S., Srinivas, K., Grace, J.K., Madhavi, M., and
649 Gajbhiye, P.N.: Soil fertility and quality assessment under tree-, crop-, and pasture-based landuse systems
650 in a rainfed environment, *Communications in Soil Science and Plant Analysis*, 40, 1436–1461, 2009.
- 651 Singh, A.K., Bordoloi, L.J., Kumar, M., Hazarika, S., Parmar, B.: Land use impact on soil quality in eastern
652 Himalayan region of India. *Environ. Monit. Assess.* 186, 2013–2024, 2014.
- 653 Smyth A.J., and Montgomery, R.F.: *Soils and landuse in central western Nigeria*. Government Printer; Ibadan,
654 Nigeria, 1962.
- 655 Soil Survey Staff.: *Keys to soil taxonomy*, 10th Ed. USDA-natural resources conservation service, Washington,
656 DC, 2006.
- 657 Sonwa D.J., Weise, S.F., Schroth, G., Janssens, M.J.J., and Shapiro, H.: Plant diversity management in cocoa
658 agroforestry systems in West and Central Africa—effects of markets and household needs, *Agroforestry*
659 *Systems*, 88, 1021–34. doi:10.1007/s10457-014-9714-5, 2014.
- 660 Tesfahunegn, G.B., Tamene, L. and Vlek, P.L.G.: Evaluation of soil quality identified by local farmers in Mai-
661 Negus catchment, northern Ethiopia. *Geoderma*. 163(3-4):209–218, 2011.
662
- 663 Tesfahunegn, G.B.: Soil quality assessment strategies for evaluating soil degradation in northern Ethiopia.
664 *Applied and Environmental Soil Science*. 2014:1–14, 2014.
665
- 666 Tesfahunegn, G.B.: Soil quality indicators response to land use and soil management systems in northern
667 Ethiopia's Catchment. *Land Degradation and Development*. 27, 438–448, 2016.
- 668 Thomaz, E.L., and Luiz, J.C.: Soil loss , soil degradation and rehabilitation in a degraded land area in Guarapuava
669 (BRAZIL), *Land Degrad. Dev.*, 23: 72–81, 2012.
- 670 Tondoh, J.E., Kouamé, F.N., Guéi, A.M., Sey, B., Koné, A.W., and Gnessougou, N.: Ecological changes induced
671 by full-sun cocoa farming in Côte d'Ivoire. *Global Ecology and Conservation*, 3, 575-595,
672 doi:10.1016/j.gecco.2015.02.007, 2015.
673
- 674 Trabaquini, K., Formaggio, R.A., and Galvão, L.S.: Changes in physical properties of soils with land use time in
675 the Brazilian savanna environment, *Land Degrad. Dev.*, 26, 397-408, 2015.

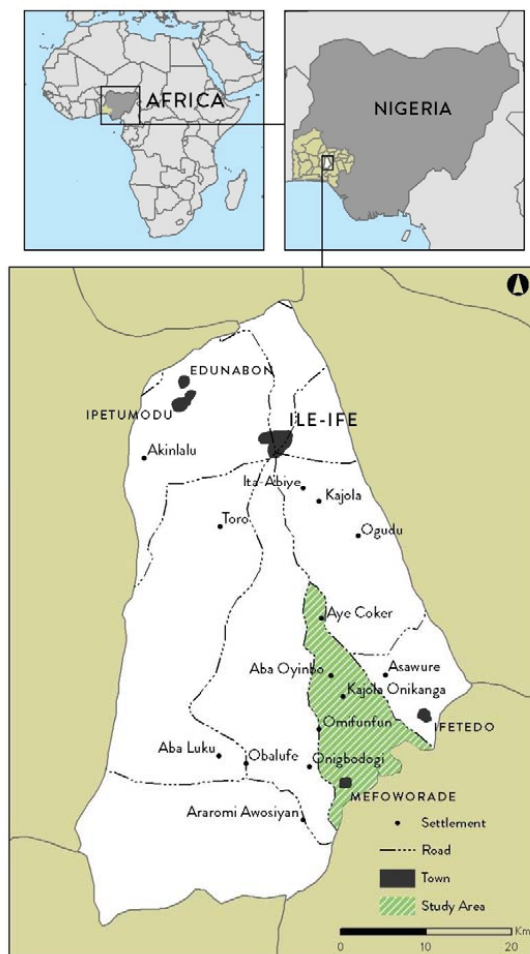


- 676
677 Virto, I., Imaz, M., Fernández-Ugalde, O., Gartzia-Bengoetxea, N., Enrique, A., and Bescansa, P.: Soil
678 degradation and soil quality in western Europe: Current situation and future perspectives. Sustainability.
679 7(1):313–365, 2014.
680
- 681 Vocomil, J.A.: Porosity. In methods of soil analysis part 1 Black CA. (ed) A.S.A Madison WI, 299-314, 1965.
- 682 Walkley, A, and Black I.A.: An examination of the Degtjareff method for determining soil organic matter and a
683 proposed modification of the chromic acid titration method, Soil Science, 37, 29-38, 1934.
- 684 Wessel, M.: Fertilizer requirement of cocoa (*Theobroma cacao L.*) in south-western Nigeria. Communication 61.
685 Department of Agriculture Resources. Royal Tropical Institute Amsterdam, 1971.
- 686 World Reference Base for Soil Resources (WRB): A framework for international classification, correlation and
687 communication, FAO Rome, 2006.
688
- 689 Yamashita, N., Ohta, S., and Hardjono, A.: Soil changes induced by Acacia mangium plantation establishment:
690 Comparison with secondary forest and Imperata cylindrica grassland soils in South Sumatra, Indonesia.
691 Forest Ecology and Management, 254 : 362–370. doi:10.1016/j.foreco.2007.08.012, 2008.
- 692 Zhao, Q., Shiliang, L., Li, D., Shikui, D., and Wang, C.: Soil degradation associated with water-level fluctuations
693 in the Manwan Reservoir, Lancang River Basin, Catena 113: 226–235, 2014.
- 694 Zornoza, R., Mataix-Solera, J., Guerrero, C., Arcenegui, V., García-Orenes, F., Mataix-Beneyto, J., and Morugán,
695 A.: Evaluation of soil quality using multiple lineal regression based on physical, chemical and biochemical
696 properties. Sci. Total Environ. 378, 233–237, 2007.
697
- 698 Zornoza, R., Mataix-Solera, J., Guerrero, C., Arcenegui, V., Mataix- Beneyto, J., and Gómez, I.: Validating the
699 effectiveness and sensitivity of two soil quality indices based on natural forest soils under Mediterranean
700 conditions, Soil Biol. Biochem., 40, 2079–2087, 2008
701
- 702 Zornoza, R., Acosta, J.A., Bastida, F., Domínguez, S.G., Toledo, D.M. and Faz, A.: Identification of sensitive
703 indicators to assess the interrelationship between soil quality, management practices and human health.
704 Soil. 1(1), 173–185, 2015.
705
706



707

Figure 1: Location map of the study area



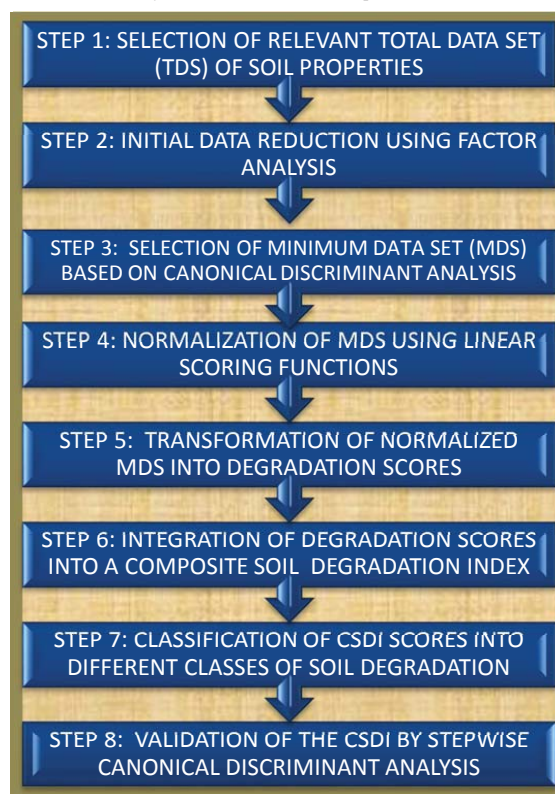
708

709



710

FIG 2. Analytical framework for development of CSDI



711
712



713

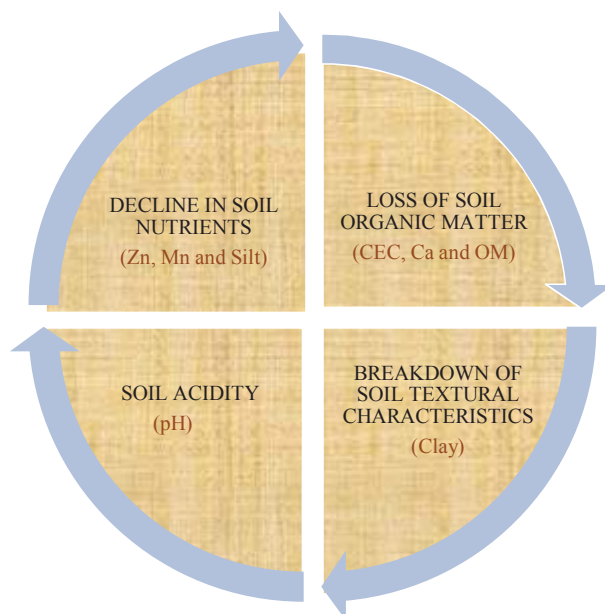
Figure 3: Scree test result from factor analysis



714
715
716
717
718



719 Fig 4. Soil degradation processes and indicators under cocoa agroforests in southwest
720



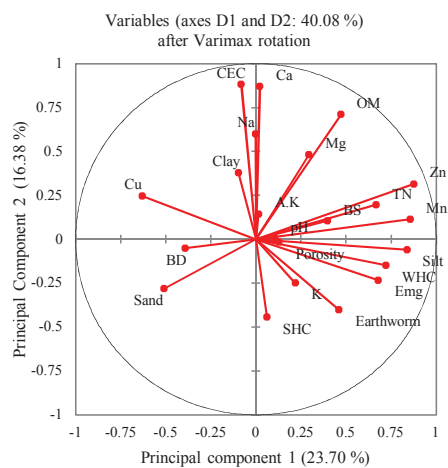


Figure 5: Principal Components' distribution of the investigated soil properties in age-sequenced peasant cocoa plantations. BD- Bulk density; WHC- Water holding capacity; SHC- Saturated hydraulic conductivity; OM- Organic matter; A.P – Available phosphorus; TN-Total nitrogen; Ca-Exchangeable calcium, Mg- Exchangeable magnesium; K- Exchangeable potassium; .Na- Exchangeable sodium; CEC- Cation exchange capacity; BS- Base saturation; Cu – Extractable copper; Zn- Extractable zinc; Mn- Extractable manganese ; EMg – Extractable magnesium; Earthworm population.

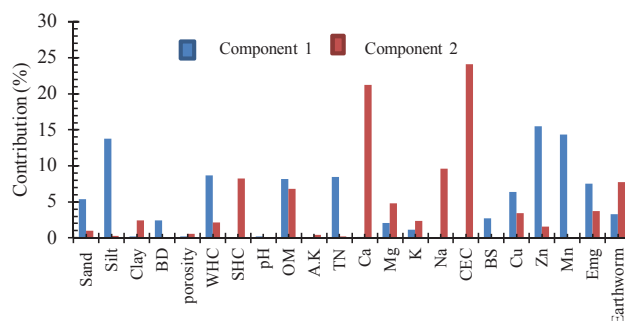


Figure 6. Percentage contributions of the investigated soil properties in age-sequenced peasant cocoa plantations. BD- Bulk density; WHC- Water holding capacity; SHC- Saturated hydraulic conductivity; OM- Organic matter; A.P – Available phosphorus; TN-Total nitrogen; Ca-Exchangeable calcium, Mg- Exchangeable magnesium; K- Exchangeable potassium; .Na- Exchangeable sodium; CEC- Cation exchange capacity; BS- Base saturation; Cu – Extractable copper; Zn- Extractable zinc; Mn- Extractable manganese ; Emg – Extractable magnesium; Earthworm population



Figure 7: First and second discriminant function separating different cocoa plantations in southwest Nigeria

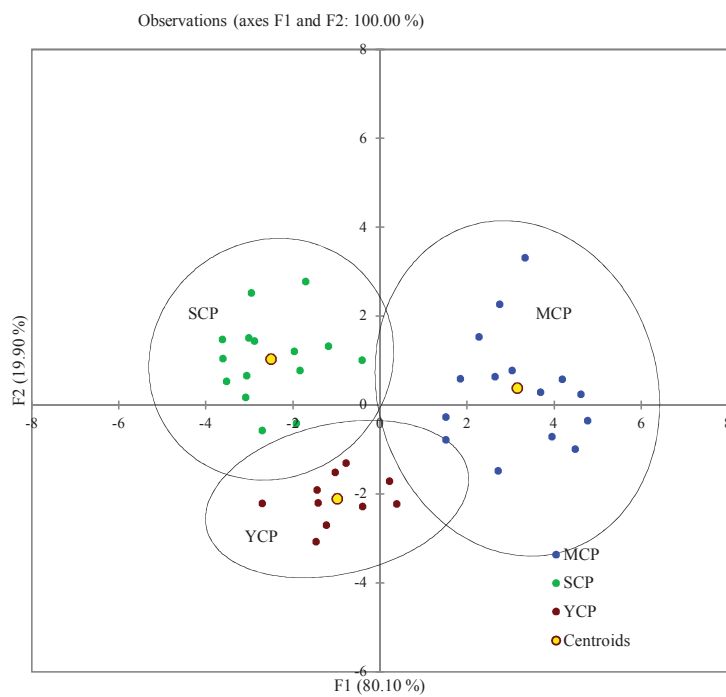




Figure 8 Percentages of degraded farms across cocoa chronosequence plantations (YCP, MCP and SCP)

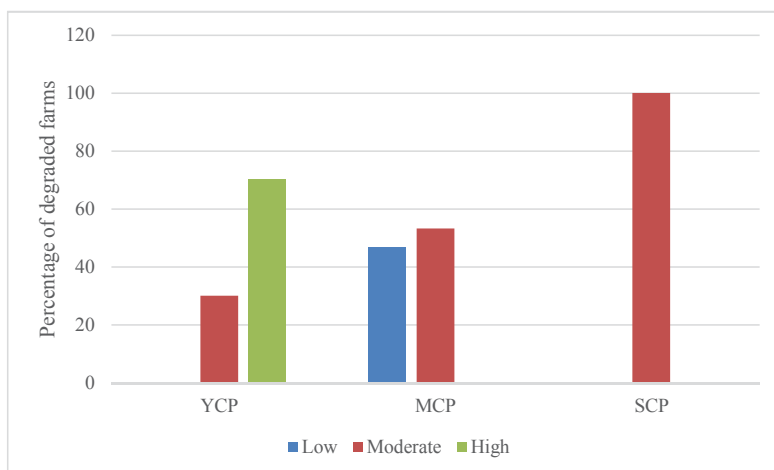




Figure 9. First and second canonical function of canonical discriminant analysis separating studied soils into three degradation classes (*Low, Moderate and High*)

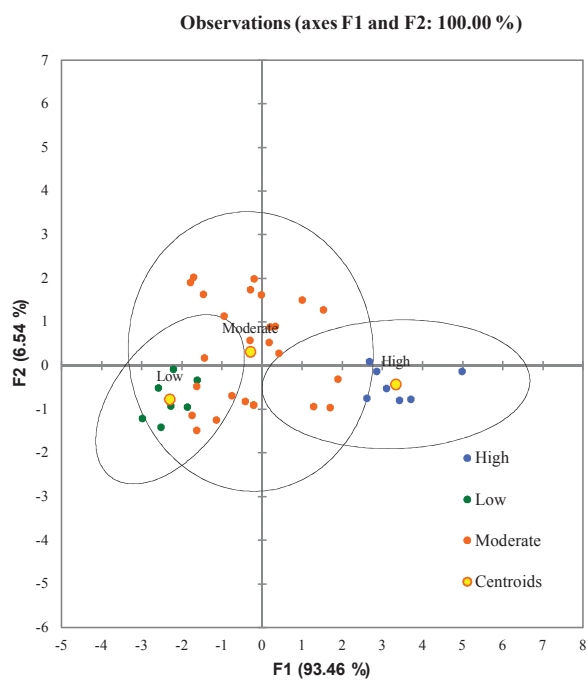




Table 1. Methods and field analysis of soil data

Soil properties	Method of determination and reference
Sand, silt and clay (%)	Pipette method (Gee & Or 2002)
Bulk density (g/cm ³).	Core method (Grossman & Reinsch 2002)
Total porosity (%)	Computed from value of bulk density (Vomocil, 1965)
Water-holding capacity (%)	Oven dry method
Saturated hydraulic conductivity (cm hr ⁻¹)	Determined in the laboratory using a constant head permeameter (Reynolds & Elrick 2002)
pH (KCl)	Potentiometrically in 0.1 M CaCl ₂ solution (Peech 1965)
Organic matter (%)	Walkley and Black (1934)
Available phosphorus (mg kg ⁻¹)	Olsen and Sommer 1982
Total nitrogen (%)	Kjeldahl method
Exchangeable Ca and Mg (mg kg ⁻¹)	Atomic absorption spectrophotometer
Exchangeable Na and K (mg kg ⁻¹)	Flame photometer
Cation exchange capacity (cmol _c kg ⁻¹)	Summation method (Juo, <i>et al.</i> 1976)
Base saturation (%)	Calculated as the percentage of the CEC occupied by basic cations
Extractable Zn, Mn, Mg and Cu (mg kg ⁻¹)	Atomic absorption spectrophotometer
Earthworm population (per m ²)	Anderson & Ingram 1993

Ca= calcium; Mg= magnesium; Na = sodium; K= potassium; Zn= zinc; Mn= manganese Cu= copper.



Table 2: Rotated factor loadings for the first five factors including proportion of variance, eigenvalues and communalities of measured soil properties

Eigenvalue	8.545	3.964	2.088	1.265	1.113	
Total Variance (%)	23.702	16.382	14.642	9.131	13.300	
Cumulative variance	23.702	40.083	54.725	63.856	77.155	
Soil degradation indicators	Principal component, PC					Communalities
	PC 1	PC 2	PC 3	PC 4	PC 5	
Sand (%)	-0.510	-0.282	-0.093	-0.094	-0.688	0.830
Extractable zinc (mg kg ⁻¹)	0.875	0.315	0.037	0.062	0.162	0.896
Extractable manganese (mg kg ⁻¹)	0.857	0.114	0.152	-0.007	0.313	0.868
Silt (%)	0.838	-0.060	-0.154	0.217	-0.014	0.777
Cation exchange capacity (cmol _c kg ⁻¹)	-0.081	0.884	-0.124	-0.094	-0.067	0.816
Exchangeable calcium (mg kg ⁻¹)	0.022	0.871	-0.007	0.028	0.084	0.767
Organic matter (%)	0.472	0.711	0.142	-0.209	0.231	0.846
Available phosphorus (mg kg ⁻¹)	0.016	0.144	0.810	0.063	0.075	0.686
Total porosity (%)	0.128	-0.016	0.801	-0.087	0.233	0.719
pH (KCl)	0.104	0.008	-0.029	0.791	0.143	0.658
Clay (%)	-0.097	0.378	0.235	-0.070	0.812	0.871
Bulk density (g cm ⁻³)	-0.393	-0.051	-0.143	-0.633	0.055	0.582
Water-holding capacity (%)	0.721	-0.147	0.358	0.367	0.278	0.882
Saturated hydraulic conductivity (cm hr ⁻¹)	0.060	-0.442	0.603	0.480	0.204	0.835
Total nitrogen (%)	0.667	0.196	0.583	0.187	0.225	0.908
Exchangeable magnesium (mg kg ⁻¹)	0.295	0.481	0.260	0.079	0.508	0.650
Exchangeable potassium (mg kg ⁻¹)	0.219	-0.249	0.099	0.094	0.624	0.518
Exchangeable sodium (mg kg ⁻¹)	-0.001	0.601	0.032	0.289	0.393	0.600
Base saturation (%)	0.397	0.104	0.355	0.272	0.661	0.806
Extractable copper (mg kg ⁻¹)	-0.632	0.247	-0.382	-0.463	-0.168	0.849
Extractable magnesium (mg kg ⁻¹)	0.679	-0.232	0.518	0.210	0.078	0.834
Earthworm population (per m ²)	0.459	-0.401	0.552	0.144	0.282	0.776

Rotation method: Varimax with Kaiser normalization.
 Boldface factor loadings are considered highly weighted;
 Extraction method: principal component analysis.



Table 3: Correlation coefficient between highly weighted variables under PC's with high factor loading

PC 1 variables	Extractable zinc	Extractable manganese	Silt
Extractable zinc	1.000	0.834**	0.653*
Extractable manganese	0.834**	1.000	0.612*
Silt	0.653*	0.612*	1.000
PC2 variables	Cation exchange capacity	Exchangeable calcium	Organic matter
Cation exchange capacity	1.000	0.870**	0.523*
Exchangeable calcium	0.870**	1.000	0.619*
Organic matter	0.523*	0.619*	1.000
PC3 variables	Available phosphorus	Total porosity	
Available phosphorus	1.000	0.578*	
Total porosity	0.578*	1.000	
PC4 variable	pH		
pH	1.000		
PC5 variable	Clay		
Clay	1.000		

* Significant difference at $P = 0.05$. ** Significant difference at $P = 0.01$.



Table 4: Result of stepwise discriminant analysis (STEPDA) separating YCP, MCP and SCP

	Discriminant function	
	1	2
Significance	0.000	0.000
Eigenvalue	6.826	1.696
% of variance	80.101	19.899
Cumulative % variance	80.101	100.000
Canonical correlation coefficient	0.934	0.793
Variables	Canonical correlation coefficients	
Silt	0.353	-0.520
Clay	0.373**	-0.139
pH	0.029	-0.211
Organic matter	0.952*	0.096
Cation exchange capacity	0.611*	0.622
Extractable Zinc	0.806*	-0.527
Available Phosphorus	0.186	-0.035
Porosity	0.158	-0.309

*, **, Significant at $p < 0.05$ and $p < 0.001$ respectively.



Table 5: CSDI value, classification and membership probabilities

CPC	CSDI Value	Z-Score value	Membership probabilities		
			Low	Moderate	High
YCP1	0.3693	0.8543	0.000	0.175	0.825
YCP2	0.3982	1.1615	0.000	0.040	0.960
YCP3	0.4421	1.6289	0.000	0.001	0.999
YCP4	0.4430	1.6379	0.000	0.001	0.999
YCP5	0.5261	2.5227	0.000	0.000	1.000
YCP6	0.3624	0.7807	0.000	0.209	0.791
YCP7	0.4238	1.4337	0.000	0.005	0.995
YCP8	0.4034	1.2173	0.000	0.030	0.970
YCP9	0.3591	0.7459	0.000	0.389	0.610
YCP10	0.3936	1.1131	0.000	0.071	0.929
MCP1	0.1916	-1.0359	0.471	0.529	0.000
MCP2	0.2175	-0.7604	0.410	0.590	0.000
MCP3	0.1977	-0.9715	0.844	0.156	0.000
MCP4	0.2333	-0.5931	0.426	0.574	0.000
MCP5	0.2386	-0.5359	0.613	0.387	0.000
MCP6	0.1757	-1.2051	0.449	0.551	0.000
MCP7	0.2790	-0.1068	0.012	0.988	0.000
MCP8	0.2669	-0.2347	0.046	0.954	0.000
MCP9	0.2584	-0.3256	0.078	0.922	0.000
MCP10	0.2564	-0.3463	0.030	0.970	0.000
MCP11	0.1187	-1.8117	0.993	0.007	0.000
MCP12	0.1836	-1.1217	0.703	0.297	0.000
MCP13	0.1645	-1.3246	0.928	0.072	0.000
MCP14	0.1476	-1.5039	0.944	0.056	0.000
MCP15	0.1367	-1.6203	0.986	0.014	0.000

CPC= Cocoa plantation chronosequence = YCP, MCP and SCP



Table 5 continue: CSDI value, classification and membership probabilities

CPC	CSDI Value	Z-SCORE value	Membership probabilities		
			Low	Moderate	High
SCP1	0.2331	-0.5948	0.100	0.900	0.000
SCP2	0.2949	0.0625	0.008	0.977	0.015
SCP3	0.2733	-0.1668	0.012	0.988	0.000
SCP4	0.2802	-0.0938	0.010	0.989	0.001
SCP5	0.3326	0.4636	0.000	0.992	0.008
SCP6	0.2851	-0.0411	0.003	0.997	0.000
SCP7	0.3242	0.3739	0.000	0.996	0.003
SCP8	0.2837	-0.0563	0.002	0.998	0.000
SCP9	0.3770	0.9365	0.000	0.995	0.005
SCP10	0.3520	0.6705	0.000	0.930	0.070
SCP11	0.2218	-0.7153	0.078	0.922	0.000
SCP12	0.2941	0.0539	0.001	0.999	0.000
SCP13	0.2589	-0.3200	0.007	0.993	0.000
SCP14	0.2918	0.0302	0.002	0.998	0.000
SCP15	0.2551	-0.3611	0.007	0.993	0.000

CPC= Cocoa plantation chronosequence = YCP, MCP and SCP



Table 6: Classification of soils into degradation levels and their interpretations modified after Gómez et al. (2009)

Range	Classes of degradation	Interpretation
< 0.195	Low	Farms with little or no form of degradation and their nutrient deficiencies can be restored with moderate effort
0.195 -0.383	Moderate	Farms with moderate soil quality degradation, where some action should be taken to improve soil conditions
> 0.383	High	Farms are currently degraded and their soil quality restoration will require sustained management efforts



Table 7: Standardized and unstandardized coefficient functions of canonical discriminant analysis

	Constant	Zn	OM	CEC	Clay
Function 1 ^ψ	-11.863	0.599*	1.225*	0.226*	0.054 ^{ns}
Function 2 ^ψ	-5.248	-0.326*	0.092 ^{ns}	0.214 ^{ns}	0.365*
Classes of degradation					
Low	-145.980	6.851	10.885	6.634	3.977
Moderate	-104.651	5.889	7.806	5.776	3.459
High	-74.970	3.359	3.489	5.202	3.564

OM- Organic matter (%); CEC- Cation Exchange Capacity (cmol_c kg⁻¹); Zn - Extractable zinc (mg kg⁻¹); Clay (%).

^ψ Wilks' lambda test of functions ($F_{\text{observed}} = 22.576$ and $F_{\text{critical}} = 2.499$) shows that the discriminant model was significant at probability $P=0.000$, for the two functions, indicating that these functions contributed more to the model.

^ψ Eigen value for $F_1 = 3.506$ and $F_2 = 0.426$;

Threshold for F_1 is $0.2/\sqrt{3.506} = 0.106$; F_2 is $0.2/\sqrt{0.426} = 0.30$

* Significant;

^{ns} Not Significant



Table 8: Cross-validation results by canonical discriminant analysis

Case	Actual group	Discriminant analysis of classification of predicted group membership					% correct
		Low	Moderate	High	Total		
Original group	from \ to						
	Low	6	1	0	7	85.71%	
	Moderate	2	23	1	26	88.46%	
	High	0	0	7	7	100.00%	
	Total	8	24	8	40	90.00%	
Cross-validated						% correct	
	from \ to	Low	Moderate	High	Total		
	Low	6	1	0	7	85.71%	
	Moderate	2	22	2	26	84.62%	
	High	0	0	7	7	100.00%	
	Total	8	23	9	40	87.50%	

Percent of "grouped" cases correctly classified = 87.50%

Boldface figure in each group is number of cases correctly classified by canonical discriminant analysis